

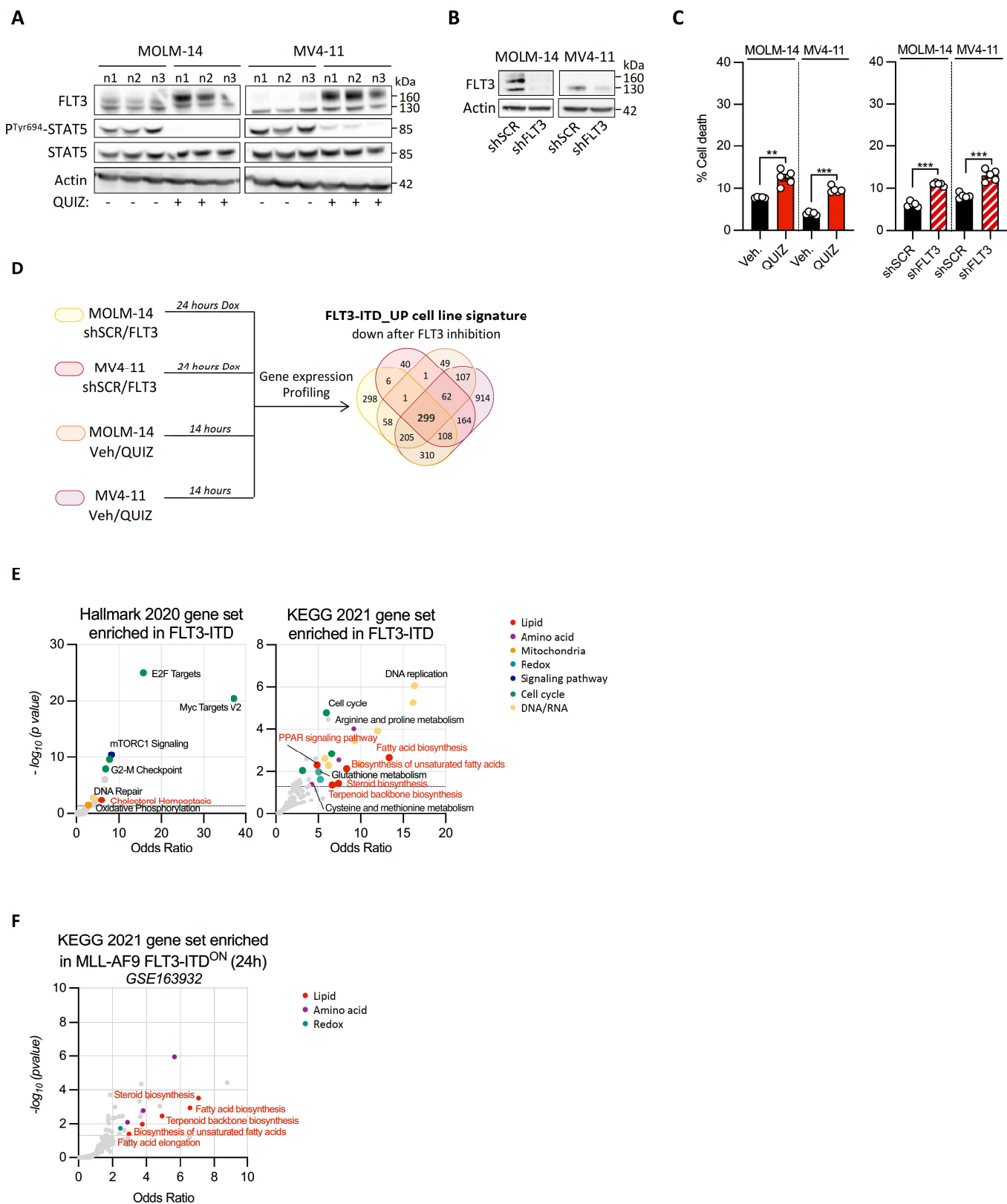


Figure S1

Supplementary Figure S1. Lipid metabolism dependency uncovered by transcriptomics in FLT3-mutant AML. **A**, Total protein extracts from MOLM-14 and MV4-11 cells incubated with vehicle or 3nM QUIZ for 14h were immunoblotted with indicated antibodies (n=3). **B**, MOLM-14 and MV4-11 cells were transduced with Dox-inducible CTL or anti-FLT3 shRNAs. Total protein extracts were immunoblotted with indicated antibodies. **C**, Cell viability was measured by PI staining in MOLM-14 and MV4-11 cells after FLT3 inhibition using 3nM QUIZ for 14h (*left panel*) or shFLT3 induction (*right panel*). **D**, Differential gene expression (DGE) was analyzed in MOLM-14 and MV4-11 cells after FLT3 inhibition using QUIZ or shFLT3, compared to vehicle or shCTL conditions, respectively. The FLT3-ITD_UP cell line signature was generated by 299 commonly downregulated genes (FC<0.67; FDR q-value <0.05) after FLT3 inhibition. **E**, Enrichr analyses of Hallmark 2020 (*left panel*) and KEGG 2021 (*right panel*) gene signatures enriched in the FLT3-ITD_UP cell line signature (p-value<0.05). **F**, Enrichr analyses of KEGG 2021 gene signatures enriched in upregulated genes in FLT3-ITD^{ON} compared to FLT3-ITD^{OFF} MLL-AF9 murine AML cells from the GSE163932 dataset (p-value<0.05).

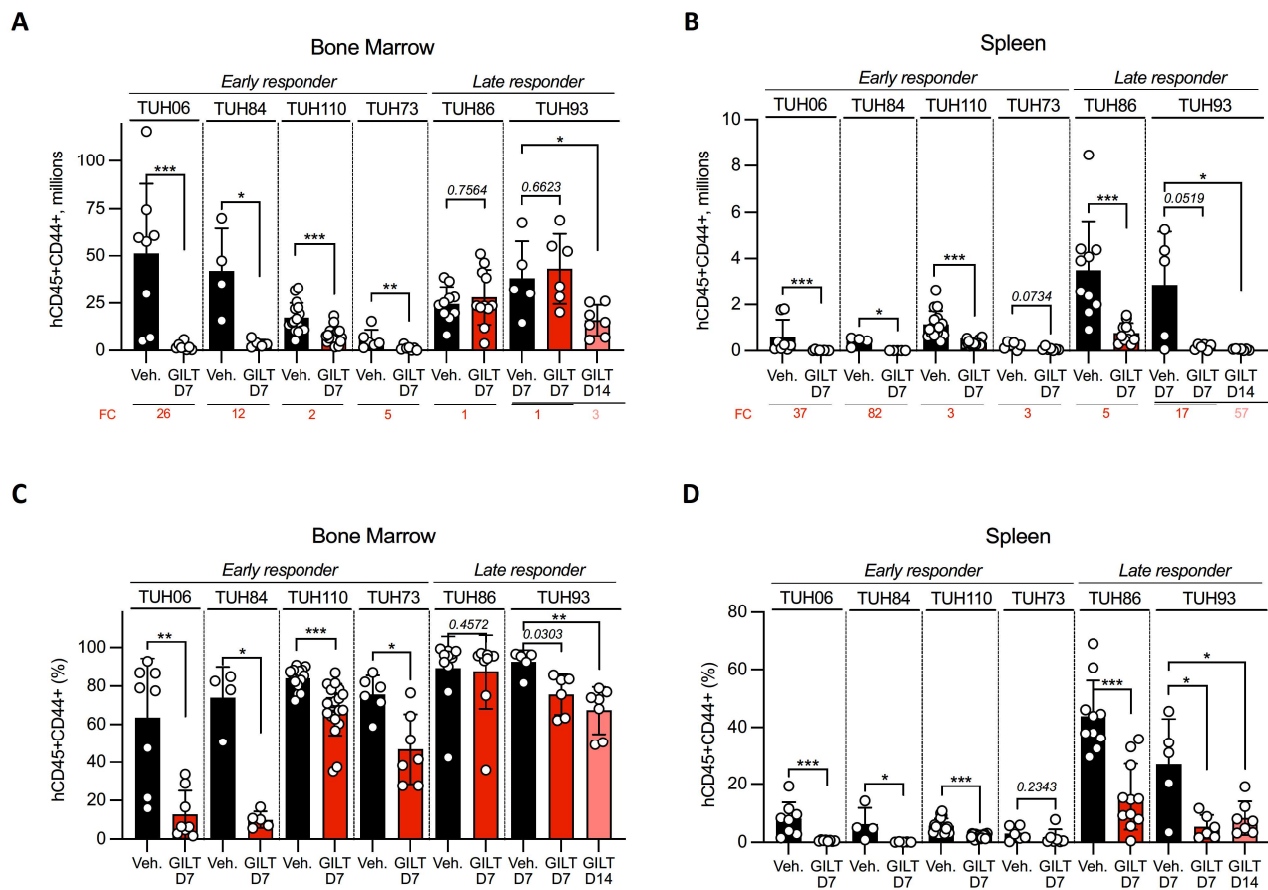
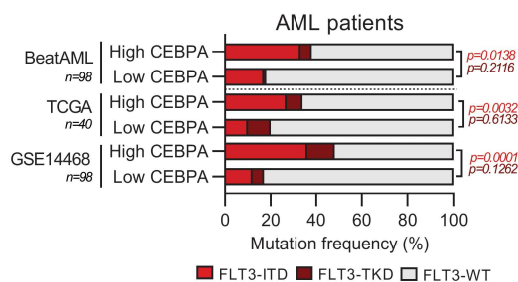


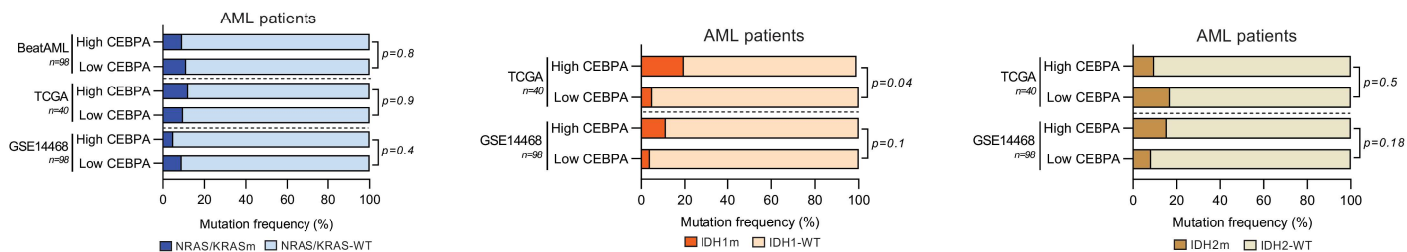
Figure S2

Supplementary Figure S2. Anti-leukemic activity of GILT across a panel of AML PDX *ex vivo* and *in vivo*. **A-B**, Number of viable CD33⁺CD45⁺ human AML cells in bone marrow (**A**) and spleen (**B**) of six AML PDXs in vehicle and GILT groups. Fold change (FC) between the mean number of leukemic cells in vehicle and GILT groups are provided. **C-D**, Percentage of viable CD33⁺CD45⁺ human AML cells in bone marrow (**C**) and spleen (**D**) of six AML PDXs in vehicle and GILT groups. Vertical bars indicate standard deviations. *: p<0.05; **: p<0.01; ***: p<0.001.

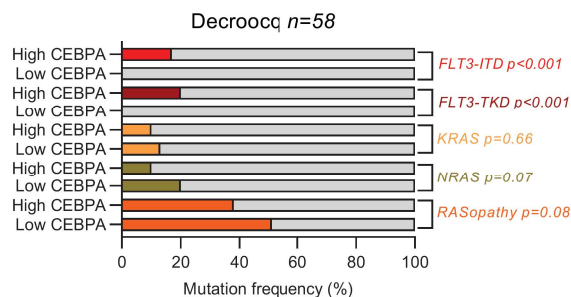
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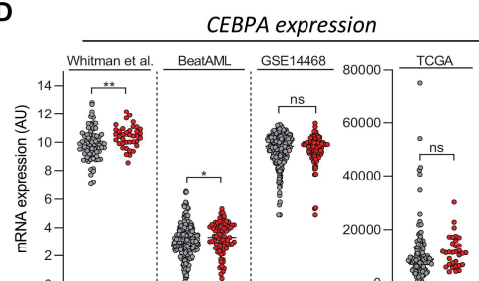
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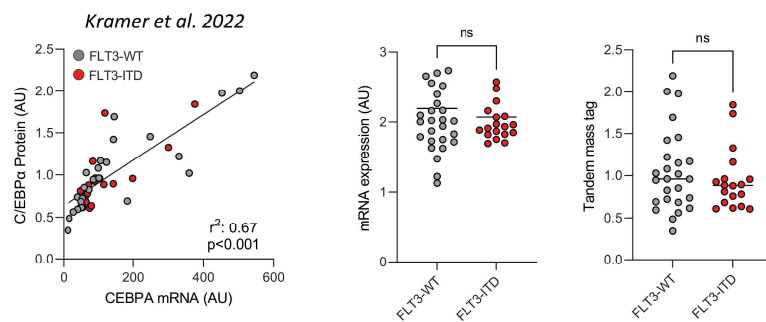
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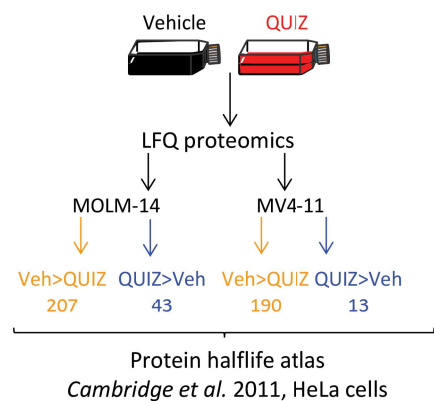
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E



F



G

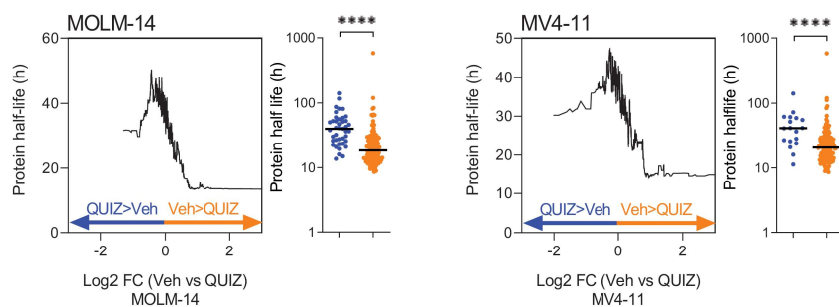
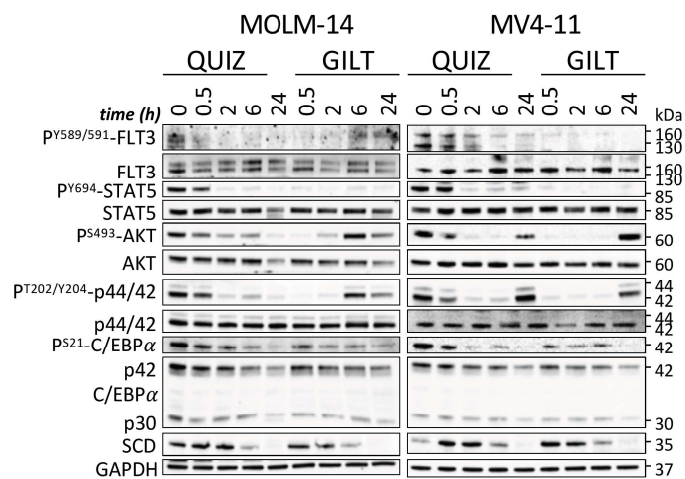


Figure S3

Supplementary Figure S3. Post-translational regulation of C/EBP α expression. **A-B**, Publicly available datasets from three independent cohorts of AML patients (BeatAML, TCGA and GSE14468) were analyzed dependent on *CEBPA* low (lower quartile) *versus* *CEBPA* high (higher quartile) expression and frequencies of *FLT3* mutation (ITD in red; TKD in dark red, **A**), *NRAS/KRAS* and *IDH1/2* (**B**). **C**, Repartition of *FLT3*, *NRAS*, *KRAS* and RASopathy mutations dependent on *CEBPA* expression in 58 AML patients from Decroocq RNAseq dataset published in (27). RASopathy referred to as *NRAS*, *KRAS*, *PTPN11* and *CBL* mutations, and *NF1* mutations and/or deletions. **D**, *CEBPA* mRNA expression in samples from patients with *FLT3* wild type (WT) or *FLT3*-ITD AML in four independent cohorts: TCGA (28), BeatAML (29), GSE14468 (30) and Whitman (31). **E**, Data from Kramer et al. (7) on 44 patients with AML including 18 *FLT3*-ITD cases were analyzed for *CEBPA* mRNA and protein (using tandem mass tag quantification) expression dependent on *FLT3* mutational status. Correlation between *CEBPA* mRNA and protein expression. Comparison of *CEBPA* mRNA (*left panel*) and protein (*right panel*) expression in *FLT3* WT *versus* *FLT3* ITD patients with AML. **F**, The half-life of proteins has been annotated from previous dataset (32). **G**, Curve represent a median half-life value calculated for each FC rank with a sliding window of size 100. Box-plot represent half-life of proteins significantly downregulated ((blue) with FC<0.77, FDR and q-value<0.05) and upregulated in Veh *versus* QUIZ conditions (FC>1.3, FDR and q-value<0.05).

A



B

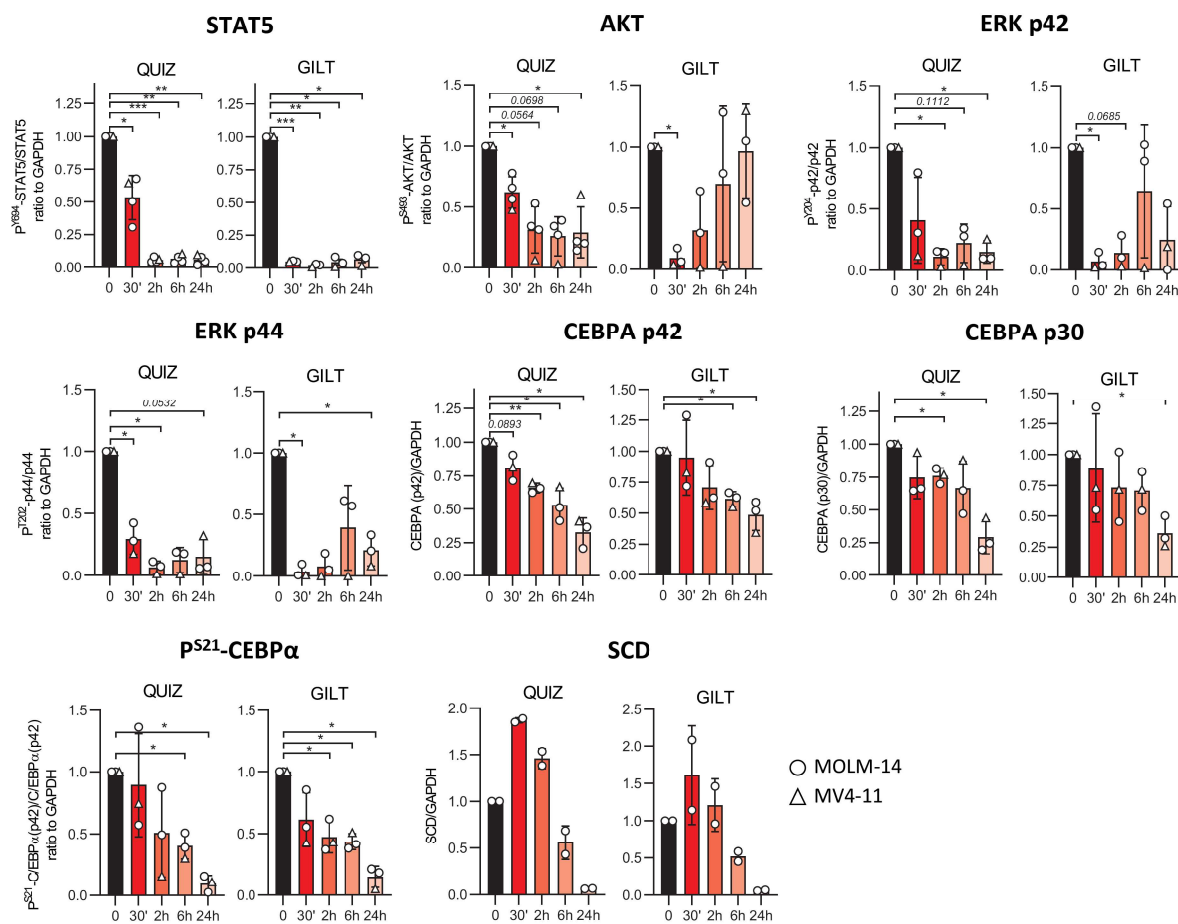
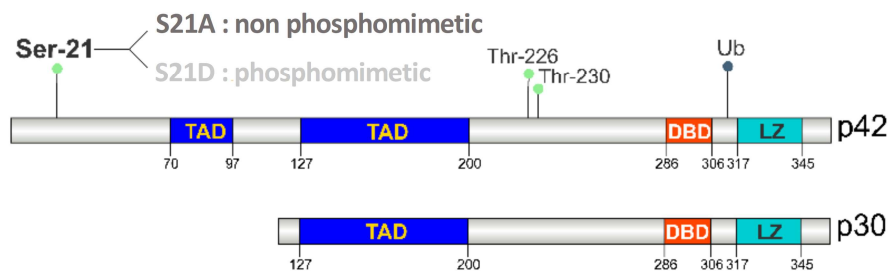
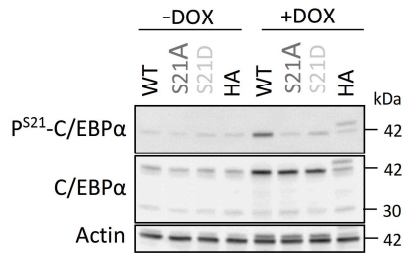
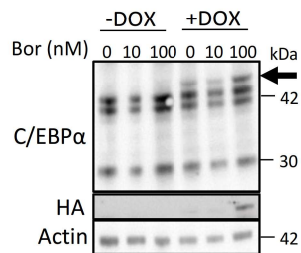
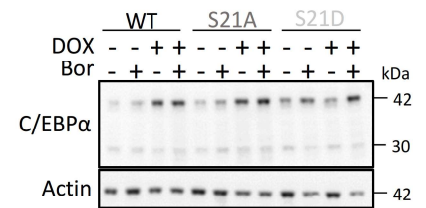
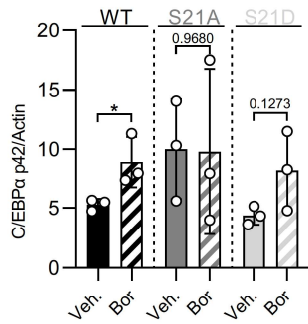


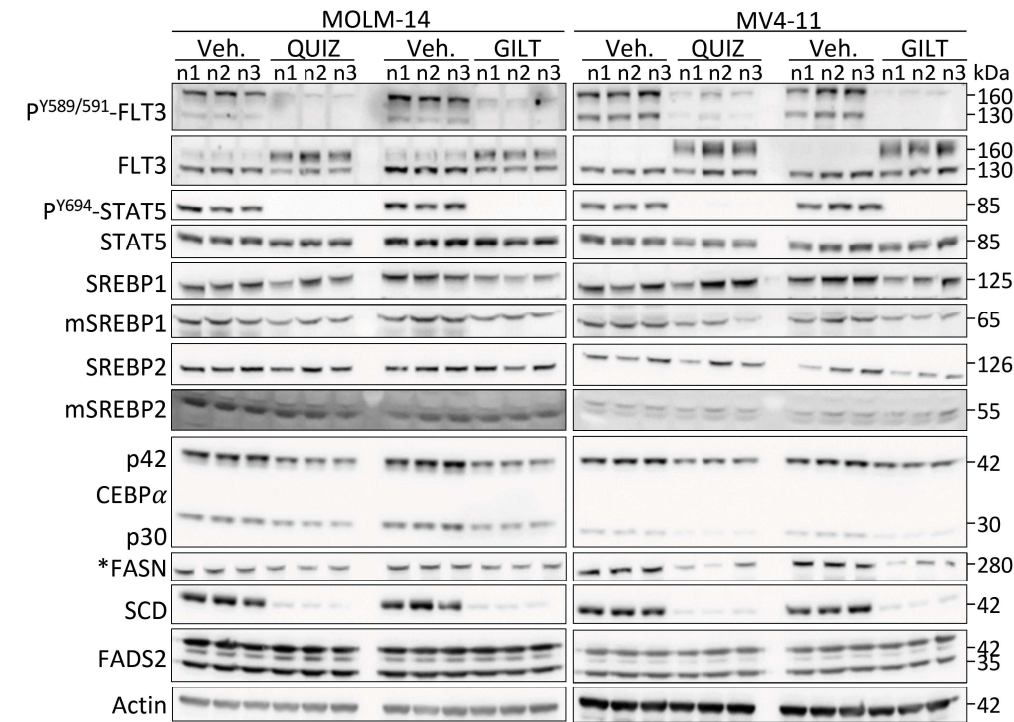
Figure S4

Supplementary Figure S4. Inhibition of C/EBP α phosphorylation and protein expression by FLT3i. **A-B**, MOLM-14 and MV4-11 cells were incubated with vehicle or 3nM QUIZ or 30nM GILT in time-course experiments as indicated. Western blots were done with the indicated antibodies (**A**) and densitometry analysis are provided (**B**). Vertical bars indicate standard deviations. *: $p < 0.05$; **: $p < 0.01$; ***: $p < 0.001$.

A**B****C****D****E****Figure S5**

Supplementary Figure S5. Ser-21 phosphorylation and post-translational regulation of C/EBP α expression. **A**, Schematic representation of C/EBP α p42 and p30 isoforms. Main domains including transcription activation domain (TAD), DNA binding domain (DBD) and leucine zipper domain (LZ) are shown, alongside with relevant post-translational modification sites. Amino acid substitutions at position 21 are shown: S21A (Serine to Alanine; non-phosphomimetic modification) and S21D (Serine to Aspartic acid; phosphomimetic change). **B-E**, MOLM-14 cells were transduced with dox-inducible expression vectors for wild type (HA-tagged or non-tagged), S21A or S21D CEBPA forms. **B**, Western blot after induction by dox using phospho-Ser-21 C/EBP α , C/EBP α and actin antibodies. **C**, CEBPA-HA cells were cultured without or with dox for 72h, and then incubated with 10nM or 100nM bortezomib (a proteasome inhibitor) for 4h. Western blot were done with C/EBP α , HA and actin antibodies. The black arrow indicates the electrophoretic migration of HA-tagged C/EBP α . **D-E**, Western blots were done on extracts from MOLM-14 CEBPA WT, S21A or S21D cells with anti-C/EBP α and anti-actin antibodies. **D**, Representative Western blot. **E**, Quantification of Western blots with a ratio between C/EBP α and actin signals (n=3). Vertical bars indicate standard deviations. ns: not significant, *p<0.05, **p<0.01.

A



B

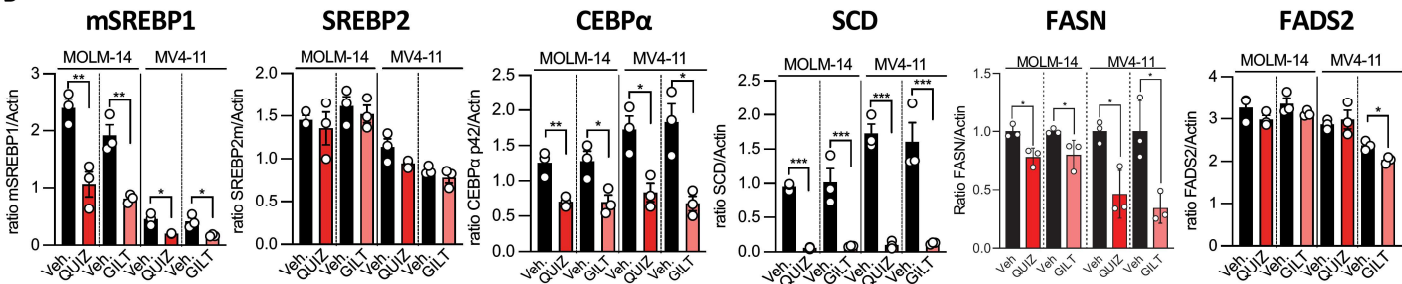


Figure S6

Supplementary Figure S6. FLT3-ITD regulates the expression of C/EBP α and of proteins related to lipid biosynthesis in AML cell lines. **A-B**, MOLM-14 and MV4-11 cells were incubated with 3nM QUIZ or 30nM GILT for 14h. Total protein extracts were submitted to Western blots with indicated antibodies (**A**). Densitometry analyses are provided (n=3) (**B**). Vertical bars indicate standard deviations. *p<0.05, **p<0.01, ***p<0.001. The *FASN symbol indicate that Western blots for FASN expression were done on different replicates (n=3) than other Western blots in this panel. FASN densitometry analysis were done relative to actin. Western blots done on these new protein extracts.

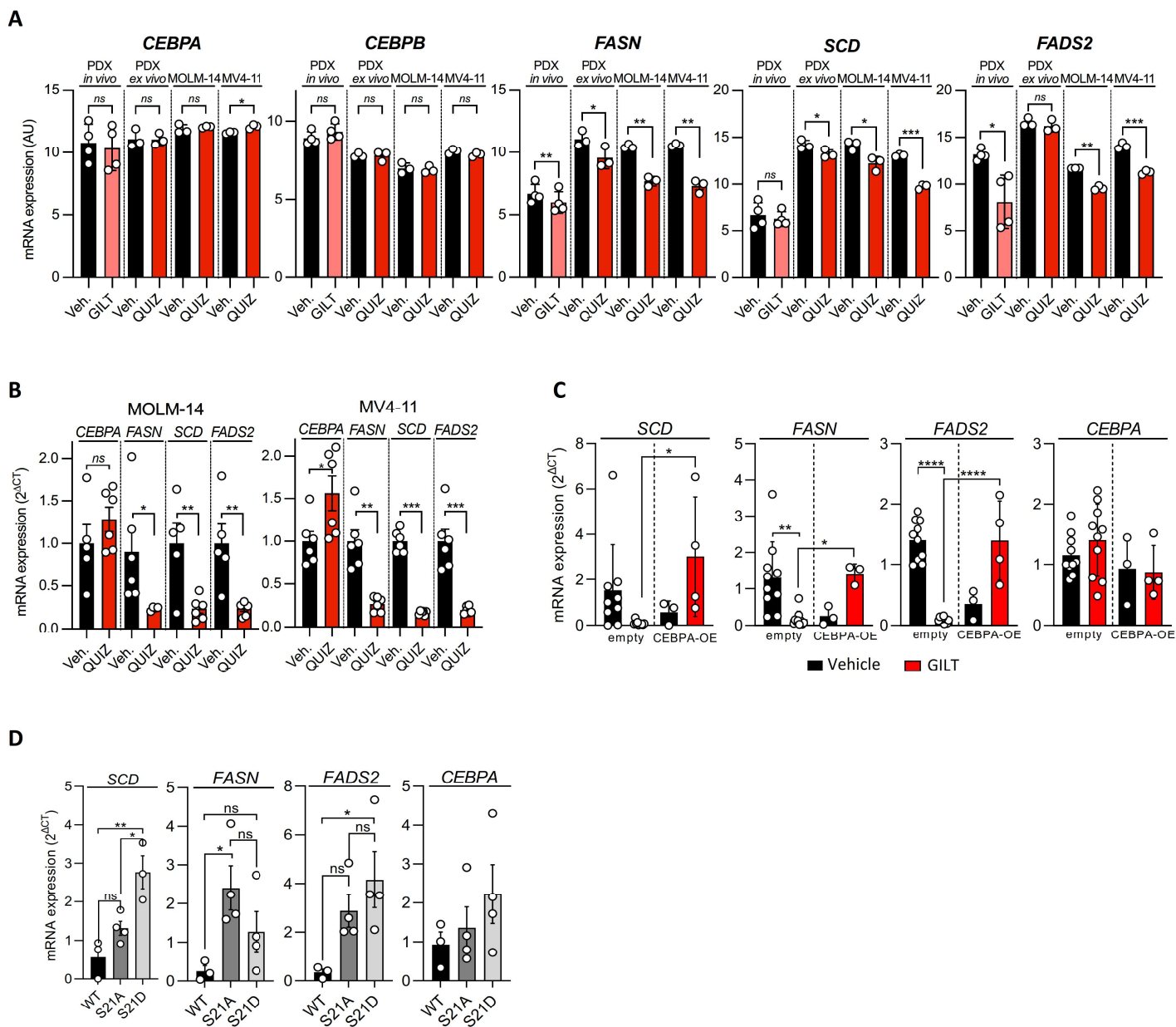
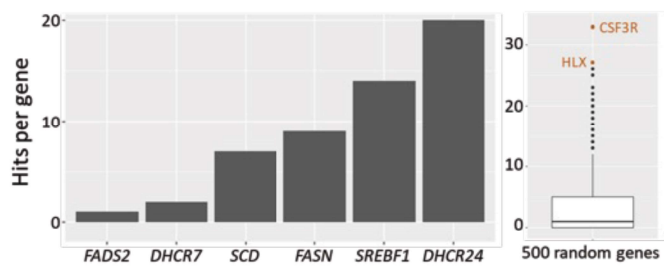
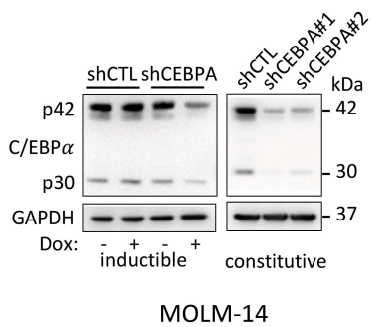
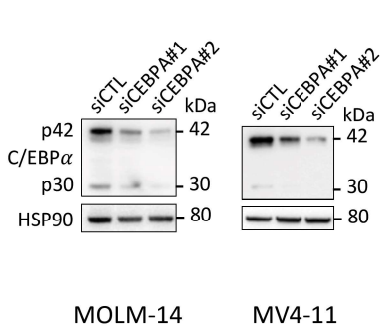
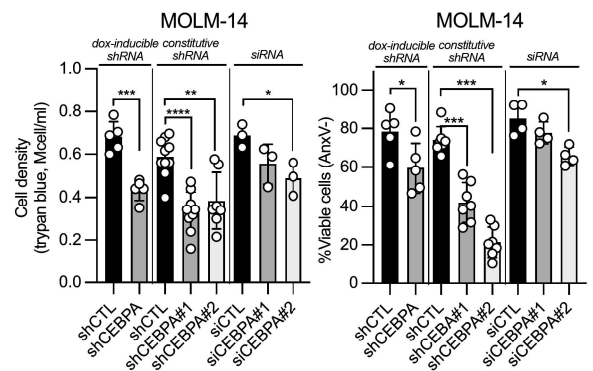
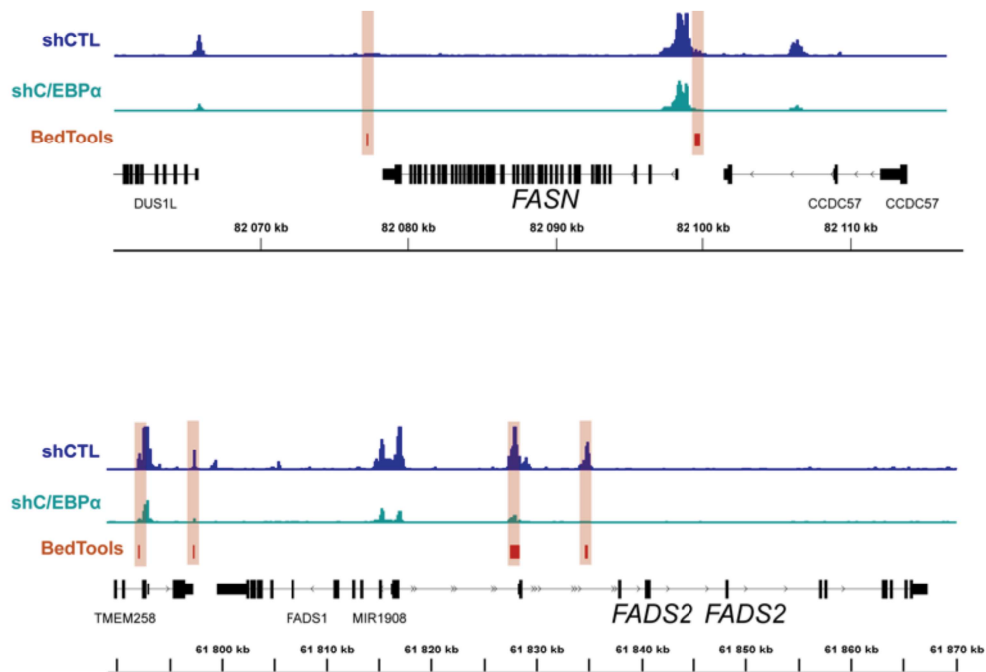


Figure S7

Supplementary Figure S7. FLT3-ITD regulates the expression of genes related to lipid biosynthesis in AML cell lines. **A**, Gene expression of *CEBPA*, *CEBPB*, *FASN*, *SCD* and *FADS2* upon FLT3i in *in vivo*-treated PDX, *ex vivo*-treated PDX and AML cell lines (MOLM-14 and MV4-11) from transcriptomic analysis. **B**, MOLM-14 and MV4-11 cells were incubated with 3nM QUIZ or 30nM GILT for 14h. Gene expression of *CEBPA*, *FASN*, *SCD* and *FADS2* upon QUIZ was investigated by quantitative RT-PCR. **C**, MOLM-14 cells transduced with empty vector (CTL) or CEBPA expression vector (CEBPA-OE) were incubated with vehicle or 30nM GILT for 14h, and mRNA expression of *SCD*, *FASN*, *FADS2* and *CEBPA* was investigated by quantitative RT-PCR. **D**, MOLM-14 cells transduced with dox-inducible WT, S21A or S21D CEBPA vectors were incubated with dox for 72h. *SCD*, *FASN*, *FADS2* and *CEBPA* mRNA expression was measured by quantitative RT-PCR. Vertical bars indicate standard deviations. *p<0.05, **p<0.01, ***p<0.001.

A**B****C****D****E****Figure S8**

Supplementary Figure S8. C/EBP α directly regulates the transcription of lipid biosynthesis genes in AML. **A**, Analysis of public C/EBP α chromatin immunoprecipitation sequencing (ChIPseq) databases including CEBPA_AML, human, AML, CEBPA (published in (33), data on MV4-11, MOLM-13, U937, SKH1, Kasumi, U937 and THP1 AML cell lines), GSE102697 (published in (34), data on SKH1 and Kasumi-1 myeloid cell lines) GSM2345026 (not published, involving MV4-11) using UniBind to map direct interactions between C/EBP α and DNA (35). C/EBP α binding on the promoter region of *SCD*, *FASN*, *FADS2*, *SREBF1*, *DHCR7* and *DHCR24*, and on the promoter region of 500 randomly chosen genes was measured as number of hits per gene. Known C/EBP α targets CSF3R and HLX are highlighted. **B-C**, MOLM-14 cells were transduced with dox-inducible control (CTL) or anti-*CEBPA* shRNAs and cultured with dox for 72h or were transfected with CTL or anti-*CEBPA* siRNA for 24h. Total protein extracts were immunoblotted with indicated antibodies. **D**, Cell viability assessed by trypan blue exclusion assay (*left panel*) and by annexin V binding assays (*right panel*). **E**, Track view of the *FASN* (*upper part*) and *FADS2* (*lower part*) locus showing ATAC-seq results for shCEBP α (green track) and shCTL (black track) MOLM14 AML cells, with regions of differential expression (BedTools intersect) depicted in red.

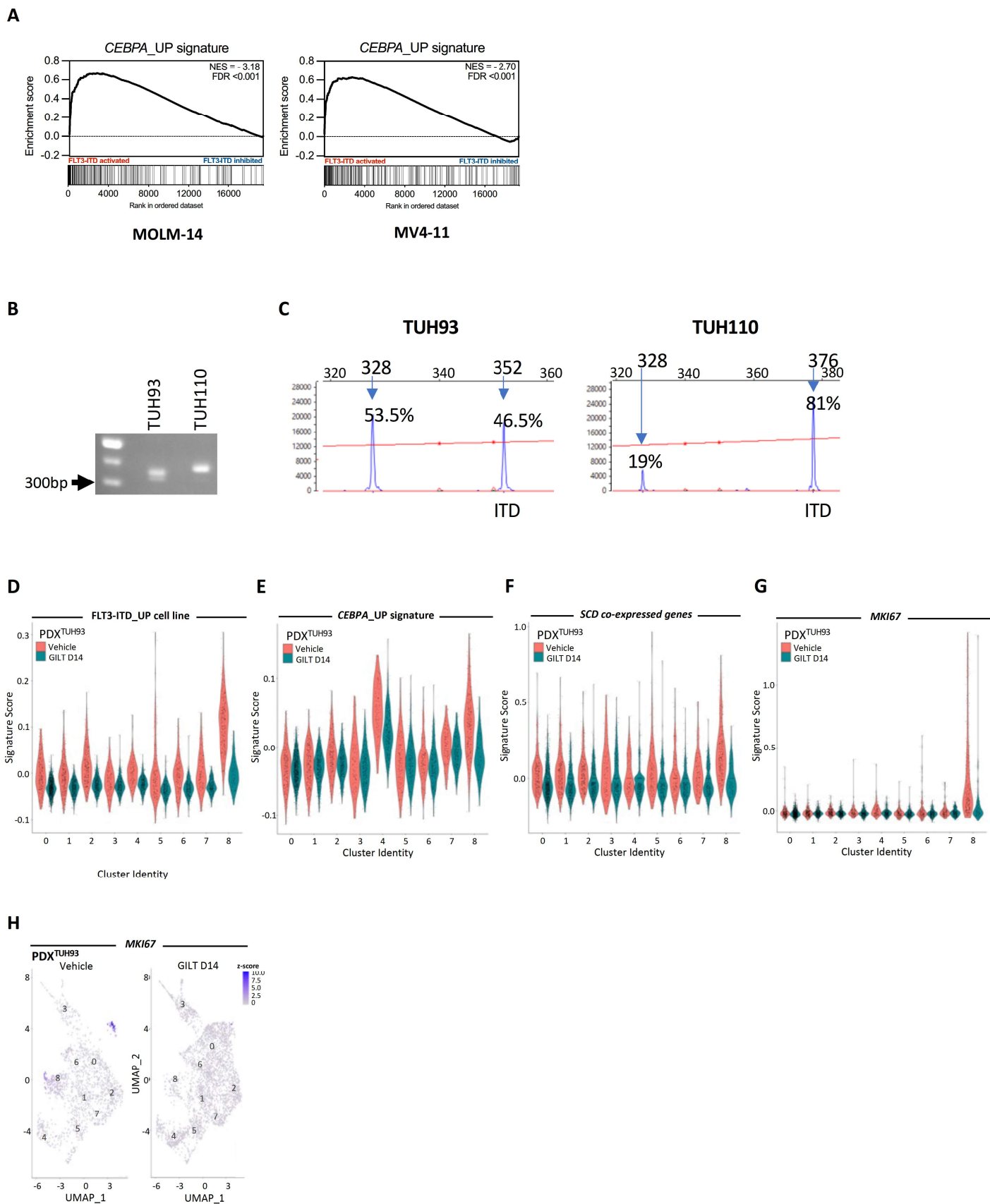


Figure S9

Supplementary Figure S9. Correlation between *CEBPA* mRNA expression and *FLT3*-ITD mutations. **A**, GSEA of *CEBPA*_UP signature in MOLM-14 (*top panel*) and MV4-11 (*bottom panel*) cells incubated with vehicle (*FLT3*-ITD activated) or 3nM QUIZ (*FLT3*-ITD inhibited) for 14h. **B-C**, Variant allele frequency of the *FLT3*-ITD mutation in PDX AML samples. Bulk DNA samples from PDX^{TUH93} and PDX^{TUH110} were extracted from purified human leukemic cells. Then the region surrounding the ITD was amplified by PCR using the following primers: 5' 6FAM-GCAATTTAGGTATGAAAGCCAGC and 3' CTTTCAGCATTTTGACGGCAACC. **B**, Migration on a 2% agarose gel of the PCR products. **C**, Fragment analysis of the PCR products. DNA size ladders are provided in the upper X-axis (in bp) and intensity is provided in Y-axis. The *FLT3*-mutation is found at +24 and +48bp in TUH93 and TUH110, respectively and the VAF of each allele is indicated in %. **D-G**, Violin plot showing the enrichment of *FLT3*-ITD_UP cell line signature (**D**), *CEBPA*_UP signature (**E**), *SCD* co-expressed genes (**F**) and *MKI67* gene (**G**) by cluster in vehicle- compared to GILT-treated PDX^{TUH93}. **H**, Visualization on the UMAP plot of *MKI67* gene enrichment in vehicle- compared to GILT-treated PDX^{TUH93}.

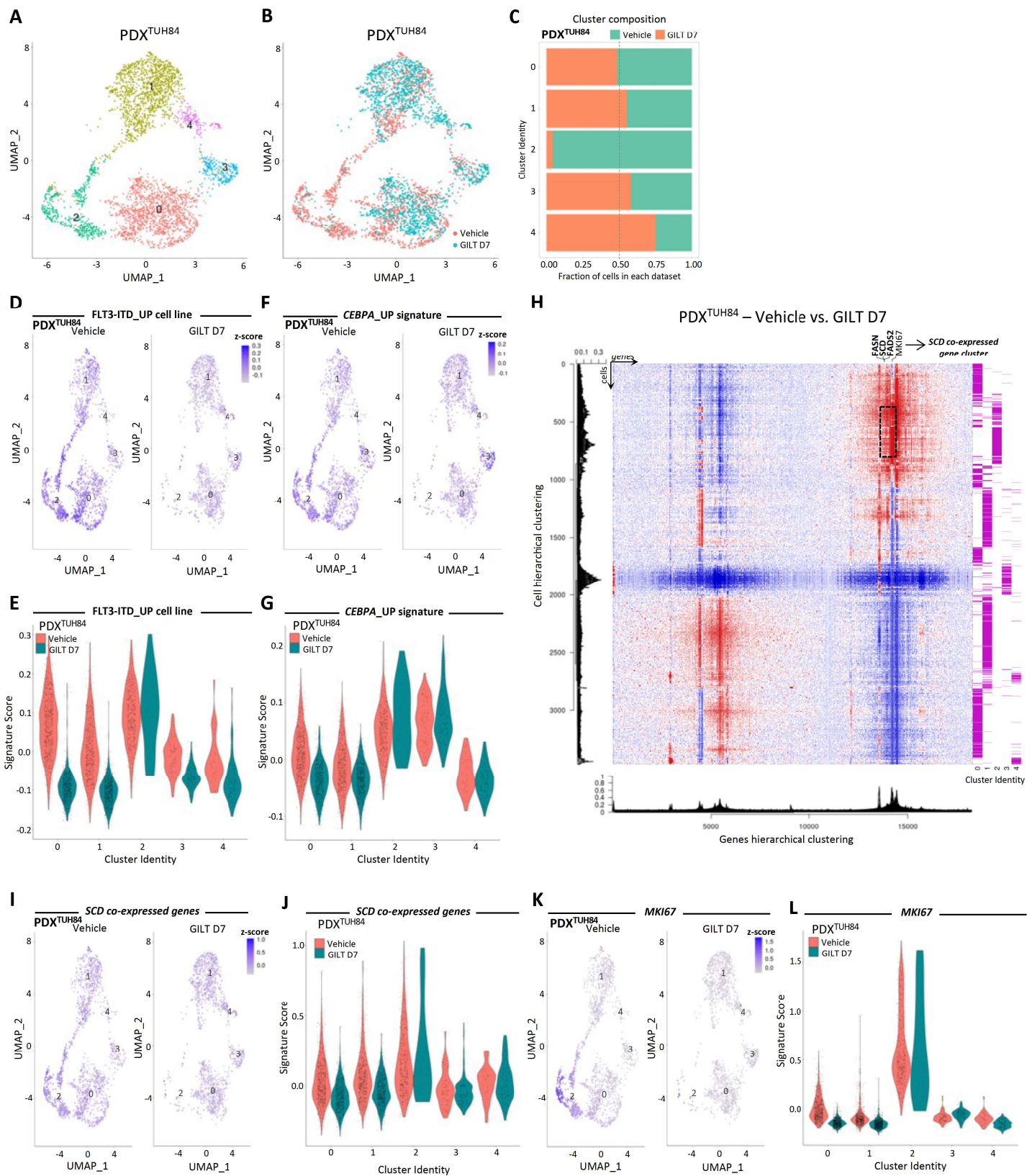


Figure S10

Supplementary Figure S10. scRNA-seq analysis from PDX^{TUH84}. **A**, Uniform manifold approximation and projection (UMAP) plot of 3623 cells from PDX^{TUH84} using Seurat. Colors indicate k-means clusters (k=8). **B**, UMAP plot colored by treatment condition (vehicle: n=1972 cells; GILT D7: n=1651 cells). **C**, Proportion of each condition per cluster from PDX^{TUH84}. **D**, Visualization on the UMAP plot of FLT3-ITD_UP cell line signature enrichment in vehicle- compared to GILT-treated PDX^{TUH84}. **E**, Violin plot showing the enrichment of FLT3-ITD_UP cell line signature by cluster in vehicle- compared to GILT-treated PDX^{TUH84}. **F**, Visualization on the UMAP plot of CEBPA_UP signature enrichment in vehicle- compared to GILT-treated PDX^{TUH84}. **G**, Violin plot showing the enrichment of CEBPA_UP signature by cluster in vehicle- compared to GILT-treated PDX^{TUH84}. **H**, Heatmap showing the expression of each gene per cell in vehicle- compared to GILT-treated PDX^{TUH84}. Dendrograms represents the hierarchical clustering of cells (*left*) and genes (*bottom*). Cluster identities are shown on the right of the Heatmap. **I**, Visualization on the UMAP plot of SCD co-expressed genes enrichment in vehicle- compared to GILT-treated PDX^{TUH84}. **J**, Violin plot showing the enrichment of SCD co-expressed genes by cluster in vehicle- compared to GILT-treated PDX^{TUH84}. **K**, Visualization on the UMAP plot of MKI67 gene enrichment in vehicle- compared to GILT-treated PDX^{TUH84}. **L**, Violin plot showing the enrichment of MKI67 gene by cluster in vehicle- compared to GILT-treated PDX^{TUH84}.

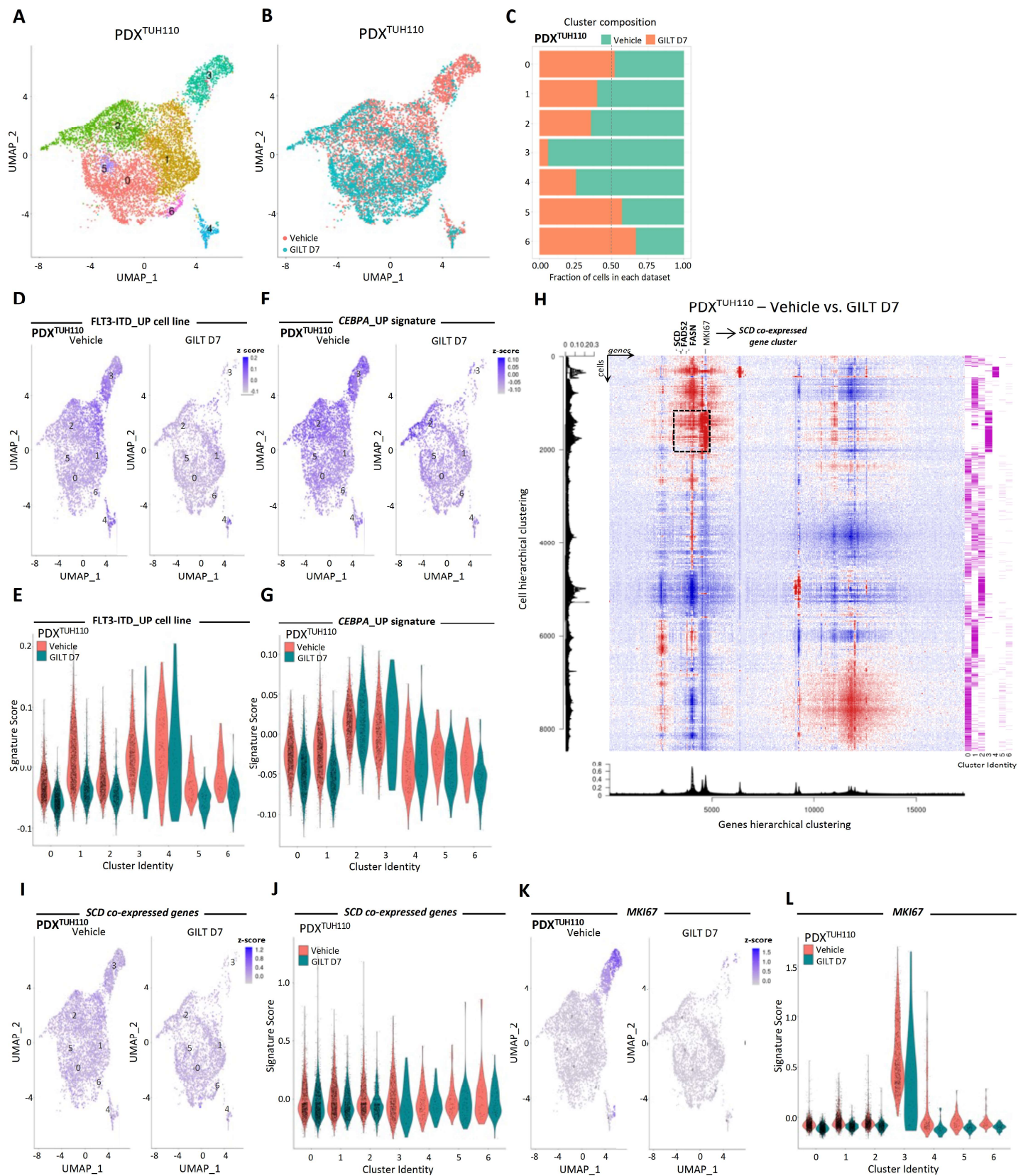


Figure S11

Supplementary Figure S11. scRNA-seq analysis from PDX^{TUH110}. **A**, Uniform manifold approximation and projection (UMAP) plot of 8739 cells from PDX^{TUH110} using Seurat. Colors indicate k-means clusters (k=8). **B**, UMAP plot colored by treatment condition (vehicle: n=5229 cells; GILT D7: n=3510 cells). **C**, Proportion of each condition per cluster from PDX^{TUH110}. **D**, Visualization on the UMAP plot of FLT3-ITD_UP cell line signature enrichment in vehicle- compared to GILT-treated PDX^{TUH110}. **E**, Violin plot showing the enrichment of FLT3-ITD_UP cell line signature by cluster in vehicle- compared to GILT-treated PDX^{TUH110}. **F**, Visualization on the UMAP plot of CEBPA_UP signature enrichment in vehicle- compared to GILT-treated PDX^{TUH110}. **G**, Violin plot showing the enrichment of CEBPA_UP signature by cluster in vehicle- compared to GILT-treated PDX^{TUH110}. **H**, Heatmap showing the expression of each gene per cell in vehicle- compared to GILT-treated PDX^{TUH110}. Dendrograms represent the hierarchical clustering of cells (*left*) and genes (*bottom*). Cluster identities are shown on the right of the Heatmap. **I**, Visualization on the UMAP plot of SCD co-expressed genes enrichment in vehicle- compared to GILT-treated PDX^{TUH110}. **J**, Violin plot showing the enrichment of SCD co-expressed genes by cluster in vehicle- compared to GILT-treated PDX^{TUH110}. **K**, Visualization on the UMAP plot of MKI67 gene enrichment in vehicle- compared to GILT-treated PDX^{TUH110}. **L**, Violin plot showing the enrichment of MKI67 gene by cluster in vehicle- compared to GILT-treated PDX^{TUH110}.

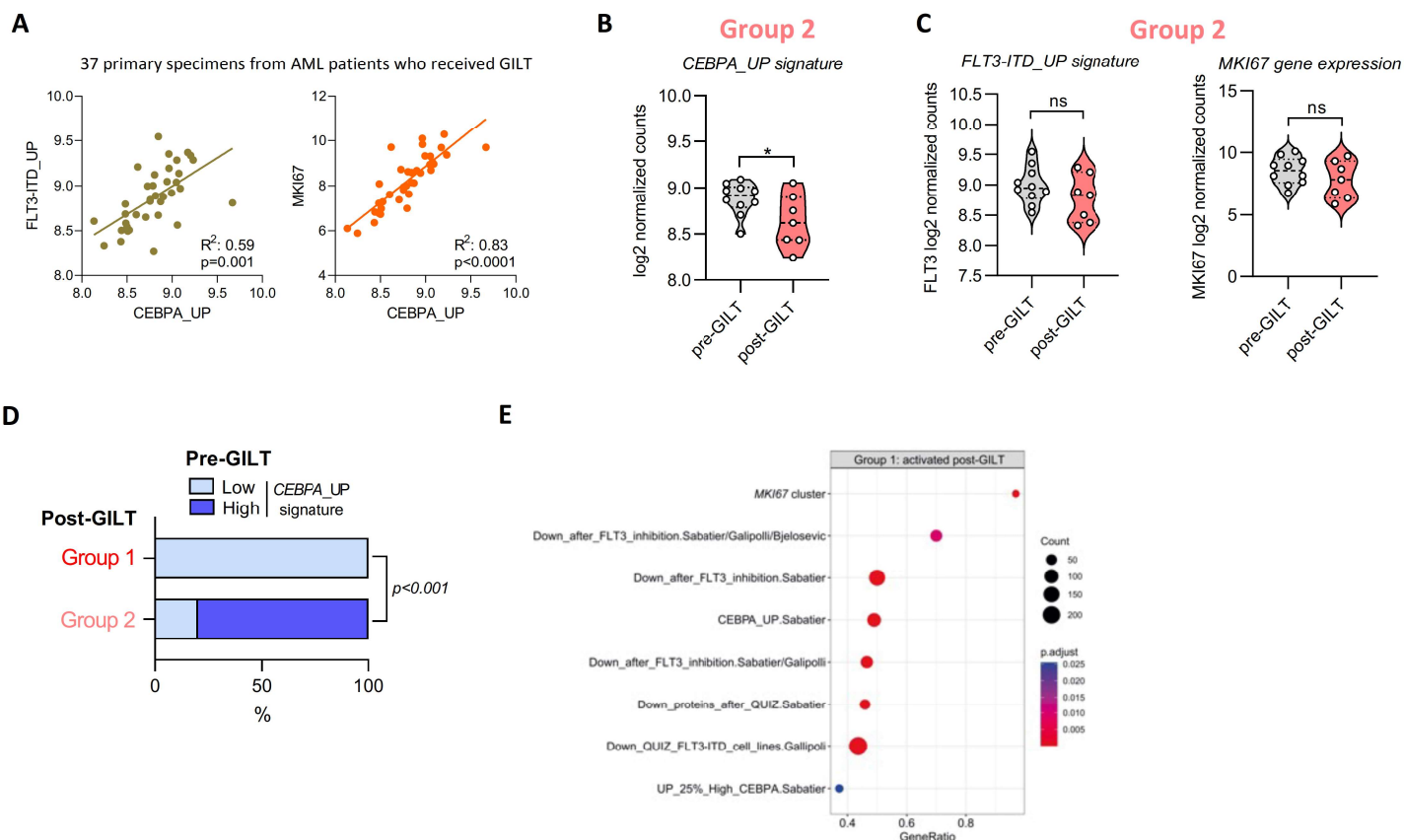


Figure S12

Supplementary Figure S12. Evolution of *CEBPA*_UP and *FLT3*-ITD_UP signatures in patients with AML treated with GILT. **A-D**, Transcriptomes from 37 samples from patients with AML (20 before and 17 after GILT treatment) were investigated. **A**, Correlation analysis for *CEBPA*_UP and *FLT3*-ITD_UP signatures and *MKI67* expression. **B-C**, Enrichment of *CEBPA*_UP, *FLT3*-ITD_UP and *MKI67* gene signatures during GILT therapy in the patient group 2. **C**, Evolution of *FLT3*-ITD_UP signature and *MKI67* expression before and after GILT treatment were plotted dependent on the post-GILT *CEBPA*_UP signature status of group 1 and 2 of AML patients. **E**, Gene set enrichment analysis comparing after *versus* before GILT in patients with enrichment in *CEBPA* signature during GILT therapy. ns: not significant, * $p < 0.05$, ** $p < 0.01$.

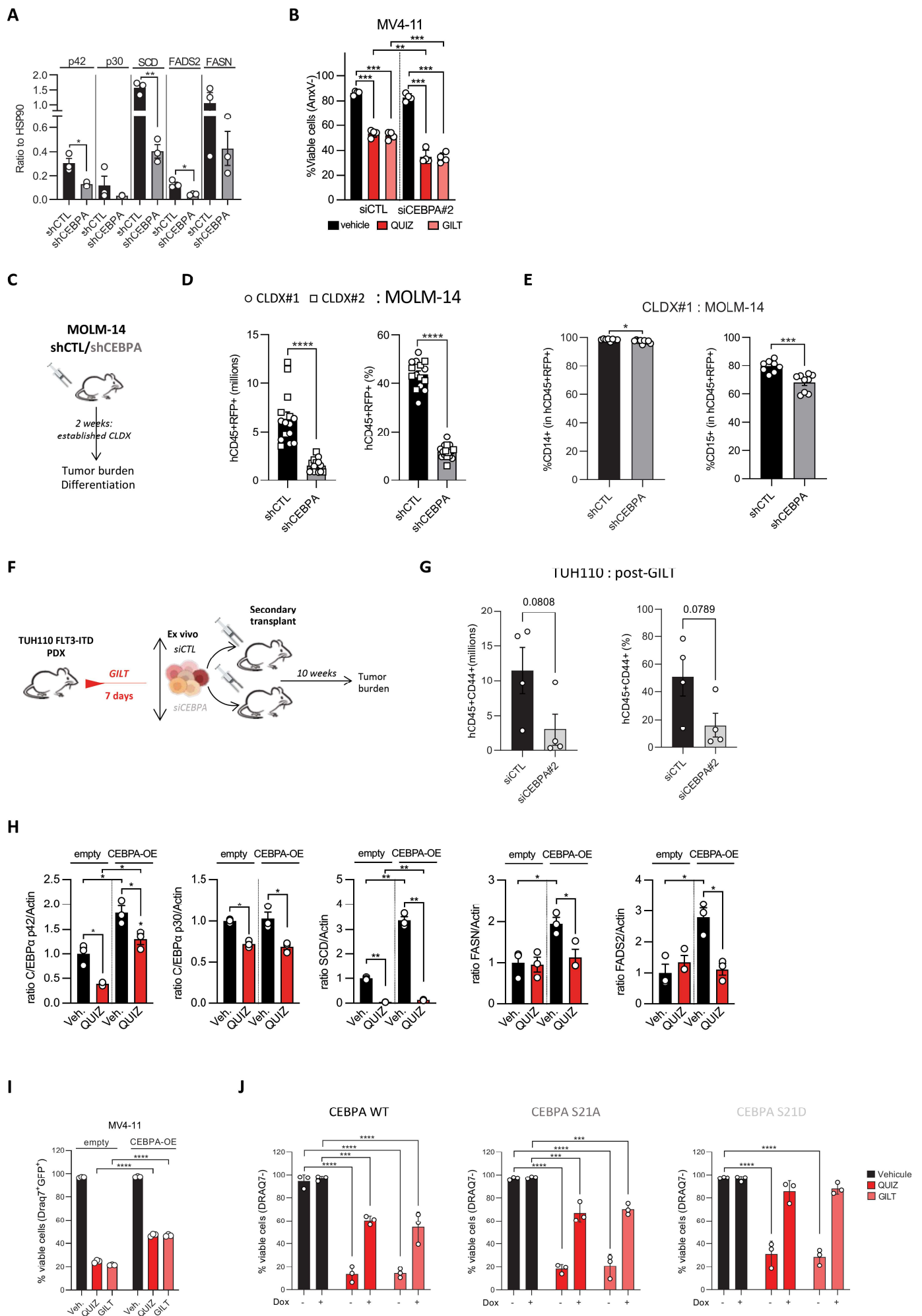


Figure S13

Supplementary Figure S13. C/EBP α regulates rate-limiting lipid biosynthetic enzymes downstream of FLT3-ITD. **A**, MOLM-14 cells transduced with dox-inducible anti-CEBPA and control (CTL) shRNAs were cultured with dox for 72h. Total protein extracts were immunoblotted with anti-CEBPA, anti-SCD and anti-actin antibodies. Densitometry analyses were done relative to actin. **B**, MV4-11 cells were transfected with anti-CEBPA (siCEBPA#2) and control (siCTL) siRNAs. Cell viability assessed by Annexin V staining after 14h incubation with 20nM QUIZ or 100nM GILT starting 6h after siRNAs transfection. **C-E**, MOLM-14 cells expressing RFP-tagged dox-inducible anti-CEBPA or CTL shRNAs were transplanted into NSG mice, and 10 days after transplant, 4mg/ml dox was added in the drinking water. Mice were euthanized 17 days after transplant. **C**, Experiment overview. **D**, Tumor burden was measured by hCD45 and RFP staining (two independent CLDX assays were performed with n=8 mice per group). **E**, Cell differentiation was investigated by CD14 and CD15 staining among hCD45⁺ human AML cells (done in CLDX#1 assay). **F-G**, PDX^{TUH110} was treated *in vivo* with 30mg/kg/d GILT for 7 days. Next, residual disease was harvested, hCD45⁺ human AML cells were sorted and transfected with CTL or anti-CEBPA siRNAs before transplantation to secondary recipient NSG mice. Tumor burden was evaluated 10 weeks after transplant. **F**, Experiment overview. **G**, Number of viable hCD45⁺CD44⁺ cells (*left panel*) and percentage of viable hCD45⁺hCD44⁺ cells (*right panel*) in bone marrow (n=4 mice per group). **H**, MOLM-14 cells transduced with a dox-inducible vector allowing CEBPA overexpression (OE), or with the empty vector were cultured with dox for 48h. Total protein extracts were immunoblotted using the indicated antibodies. Densitometry analyses were done relative to actin. **I**, MV4-11 cells transduced with CEBPA-OE or empty vector were incubated with dox for 72h, and then incubated with vehicle or 3nM QUIZ or 30nM GILT for 48h. Viability was measured by DRAQ7 staining in efficiently transduced GFP⁺ cells. **J**, MOLM-14 cells were transduced with dox-inducible expression vectors for wild type (WT), S21A and S21D CEBPA forms. After 72h dox induction, cells were incubated with vehicle, 3nM QUIZ or 30nM GILT for 48h and viability was measured using DRAQ7 staining. Vertical bars indicate standard deviations. ns: not significant; *: p<0.05; **: p<0.01; ***: p<0.001.

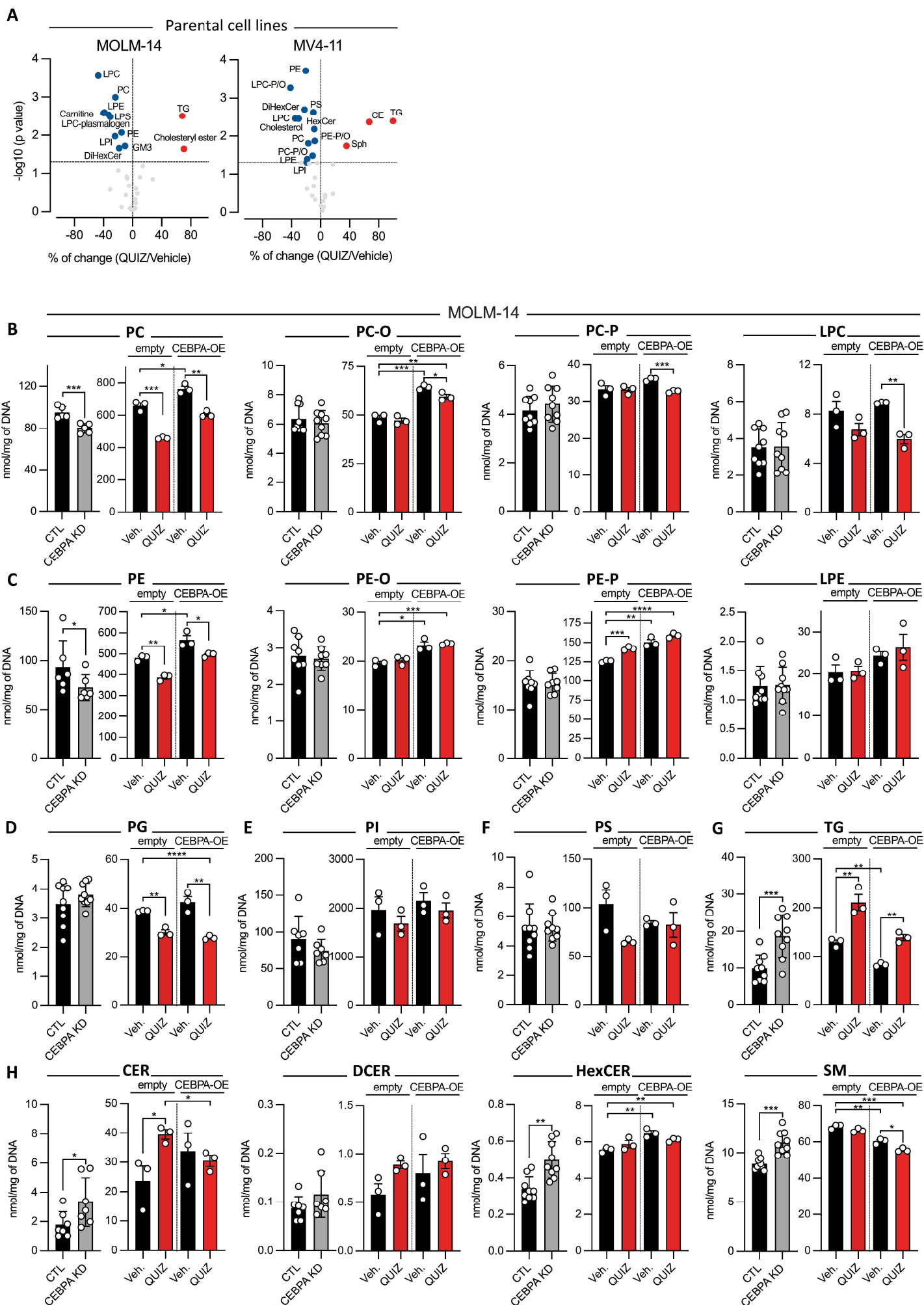


Figure S14

Supplementary Figure S14. C/EBP α controls lipid amount in FLT3-ITD cell lines. **A**, Global lipidomics in MOLM-14 (*left panel*) and MV4-11 (*right panel*) parental AML cell lines incubated with vehicle or 3nM QUIZ for 14h. LPC/PC: lyso-/phosphatidylcholine; LPE/PE: lyso-/phosphatidylethanolamine; LPI/PI: lyso-/phosphatidylinositol; LPS/PS: lyso-/phosphatidylserine; TG: triglyceride; CE/Cer: ceramide; DiHexCer: dihexosylceramide; HexCER: Hexosylceramide; SM/Sp: sphingomyelin; GM3: monosialodihexosylganglioside; -P: plasmalogen; -O: ether. **B-H**, Targeted lipidomics was performed in MOLM-14 cells were depleted from C/EBP α (siRNA 24h, dox-inducible shRNA or constitutive shRNA 72h) or overexpressing C/EBP α (dox-inducible 48h) and treated with vehicle or 3nM QUIZ for 24h. Each graph represents the quantity of each lipid species relative to DNA amount per sample. Vertical bars indicate standard deviations. *: p<0.05; **: p<0.01; ***: p<0.001.

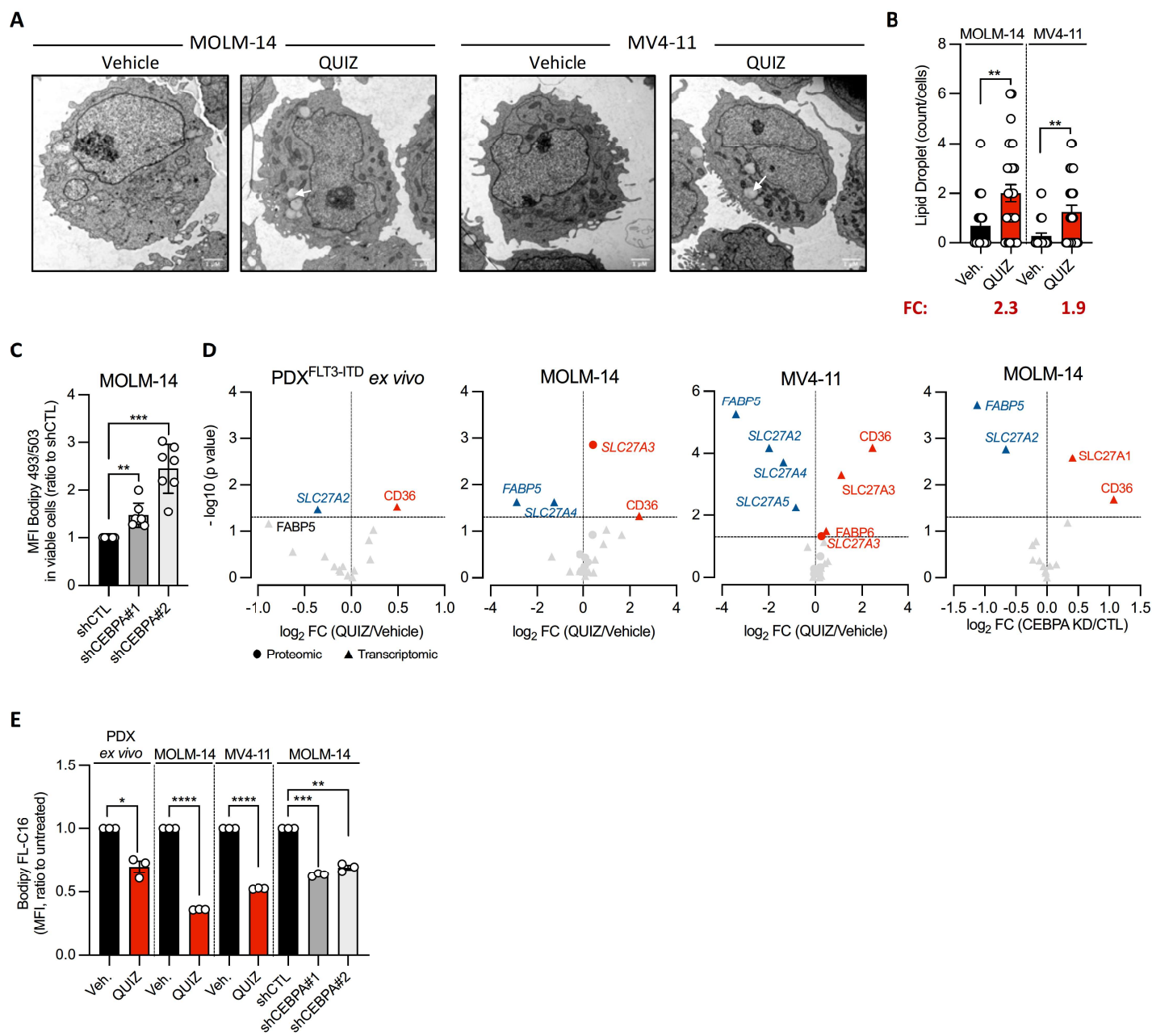


Figure S15

Supplementary Figure S15. FLT3 inhibitors induce a lipid switch increasing neutral lipids dependent on C/EBP α . **A-B**, MOLM-14 and MV4-11 cells were incubated with vehicle or 3nM QUIZ for 14h. **A**, Electron microscopy (EM) morphology (scale: 5 μ M). **B**, EM quantification of lipid droplet content *per cell*. Each dot corresponds to a single cell. **C**, Neutral lipid content in MOLM14 silenced for C/EBP α compared to control using constitutive shRNA measured by Bodipy 493/503 staining. **D**, Gene and protein expression of FA binding protein and lipid transporter in FLT3-ITD patient-derived AML cells and cell lines incubated with vehicle or QUIZ, and in MOLM-14 cells transduced with CTL or anti-CEBPA shRNAs (decrease and increase FC<0.67 and >1.5, respectively). **E**, Patient-derived FLT3-ITD AML cells and cell lines were incubated with 3nM QUIZ for 14h, and MOLM-14 cells were transduced with CTL or two different anti-CEBPA shRNAs. Lipid transport was assessed using Bodipy FL-C16 staining. Vertical bars indicate standard deviations. *: p<0.05; **: p<0.01; ***: p<0.001.

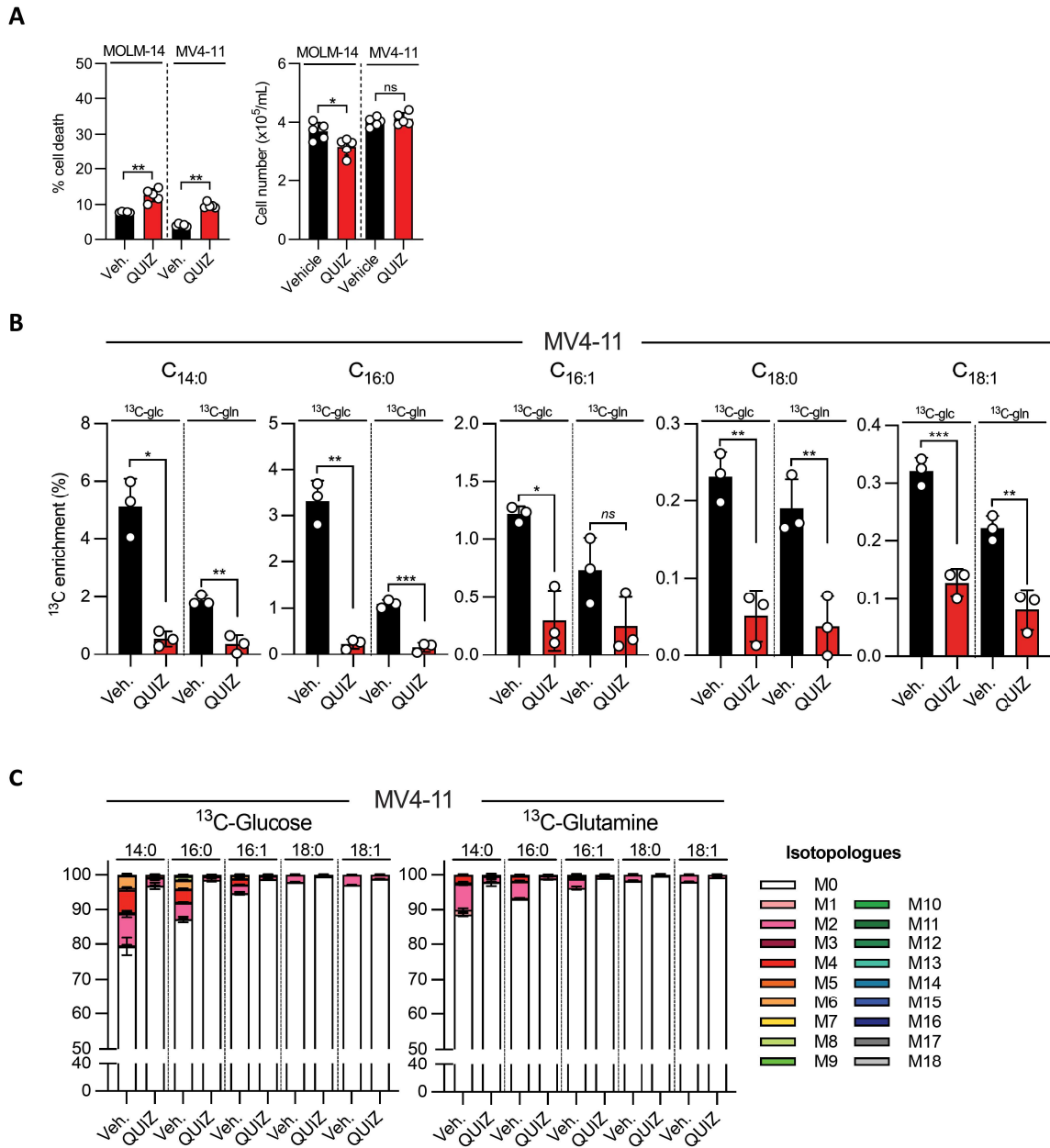


Figure S16

Supplementary Figure S16. FLT3 inhibitor inhibits fatty acid synthesis fueled by glucose and glutamine. **A**, MOLM-14 and MV4-11 cells were incubated with vehicle or 3nM QUIZ for 14h. Viability was measured by propidium iodine staining. Viable cells were also counted using a trypan blue exclusion assay. **B**, Percentage of ^{13}C enrichment in $\text{C}_{14:0}$, $\text{C}_{16:0}$, $\text{C}_{16:1}$, $\text{C}_{18:0}$ and $\text{C}_{18:1}$ total FA in MV4-11 cultured on $[\text{U-}^{13}\text{C}]$ -glucose or $[\text{U-}^{13}\text{C}]$ -glutamine and treated with vehicle or 3nM QUIZ for 24h. **C**, Isotopologues distribution in $\text{C}_{14:0}$, $\text{C}_{16:0}$, $\text{C}_{16:1}$, $\text{C}_{18:0}$ and $\text{C}_{18:1}$ total FA in MV4-11 24-cultured on $[\text{U-}^{13}\text{C}]$ -glucose (*left panel*) or $[\text{U-}^{13}\text{C}]$ -glutamine (*right panel*) and incubated with vehicle or 3nM QUIZ. Vertical bars indicate standard deviations. *: $p<0.05$; **: $p<0.01$; ***: $p<0.001$.

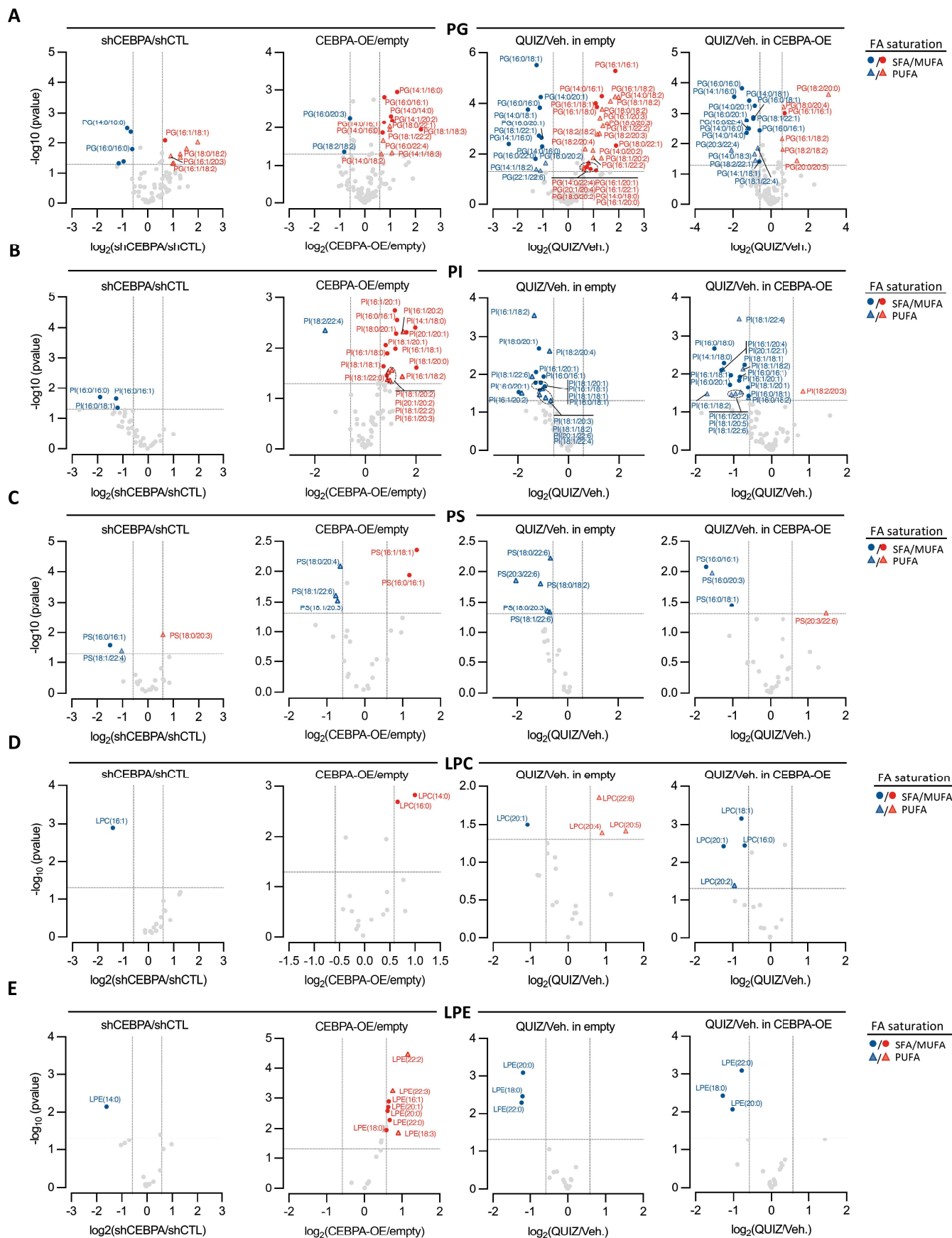


Figure S17

Supplementary Figure S17. FLT3 inhibition decreases monounsaturated fatty acid dependent on C/EBP α . **A-E**, From the *left* to the *right*: C/EBP α -depleted versus CTL, CEBPA-OE versus empty vector, vehicle versus 3nM QUIZ for 14h in empty vector, and vehicle versus 3nM QUIZ for 14h in CEBPA-OE in MOLM-14 cells. Targeted lipidomics showing repartition of SFAs, MUFAs and PUFAs distribution into phosphatidylglycerol (PG, **A**), phosphatidylinositol (PI, **B**), phosphatidylserine (PS, **C**), lysophosphatidylcholine (LPC, **D**) and lysophosphatidylethanolamine (LPE, **E**).

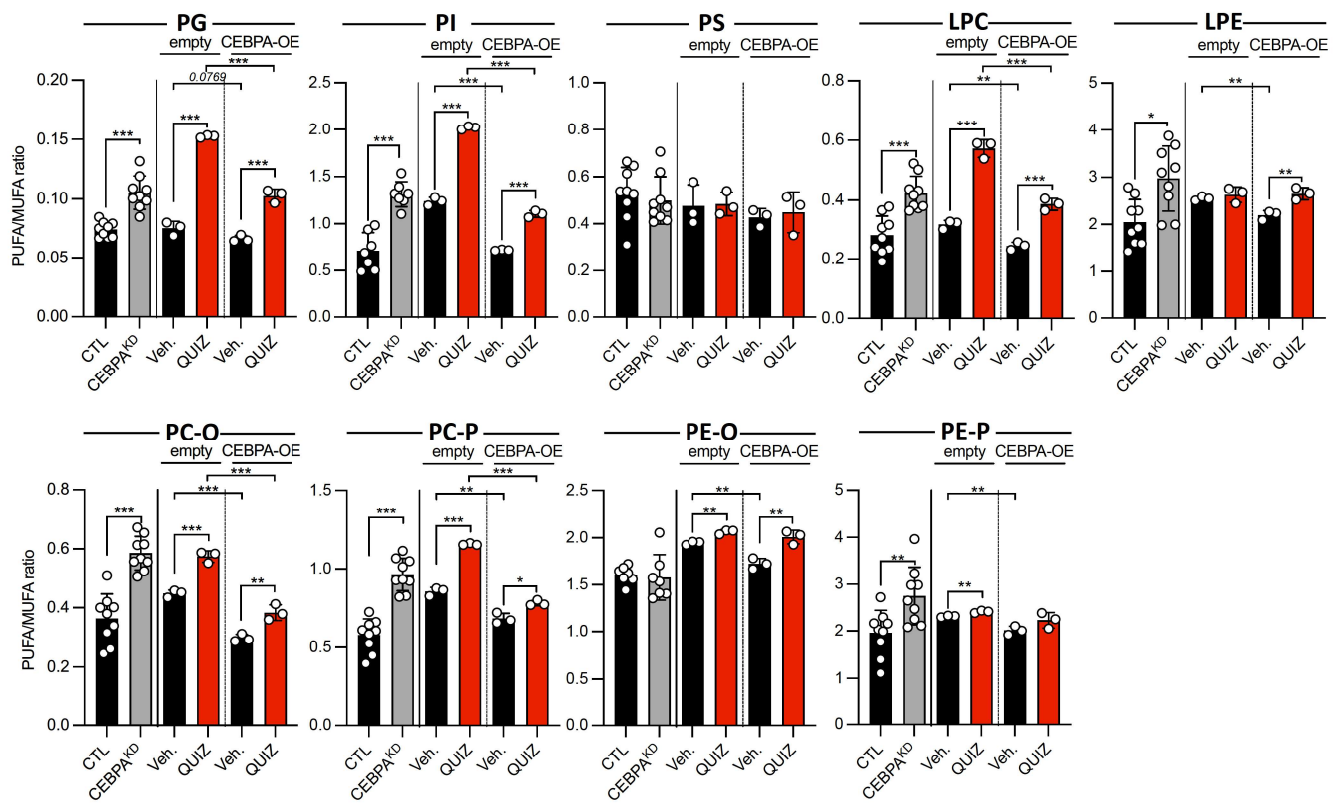


Figure S18

Supplementary Figure S18. FLT3 inhibition increases PUFA/MUFA ratio dependent on C/EBP α . From the *left* to the *right*: C/EBP α -depleted versus CTL, CEBPA-OE versus empty vector, vehicle versus 3nM QUIZ for 14h in empty vector, and vehicle versus 3nM QUIZ for 14h in CEBPA-OE in MOLM-14 cells. PUFA/MUFA ratio into phosphatidylglycerol (PG), phosphatidylinositol (PI), phosphatidylserine (PS), lysophosphatidylcholine (LPC), lysophosphatidylethanolamine (LPE), phosphatidylcholine-ether/-plasmogen (PC-O/PC-P) and phosphatidylethanolamine-ether/-plasmogen (PE-O/PE-P). Vertical bars indicate standard deviations. *: p<0.05; **: p<0.01; ***: p<0.001.

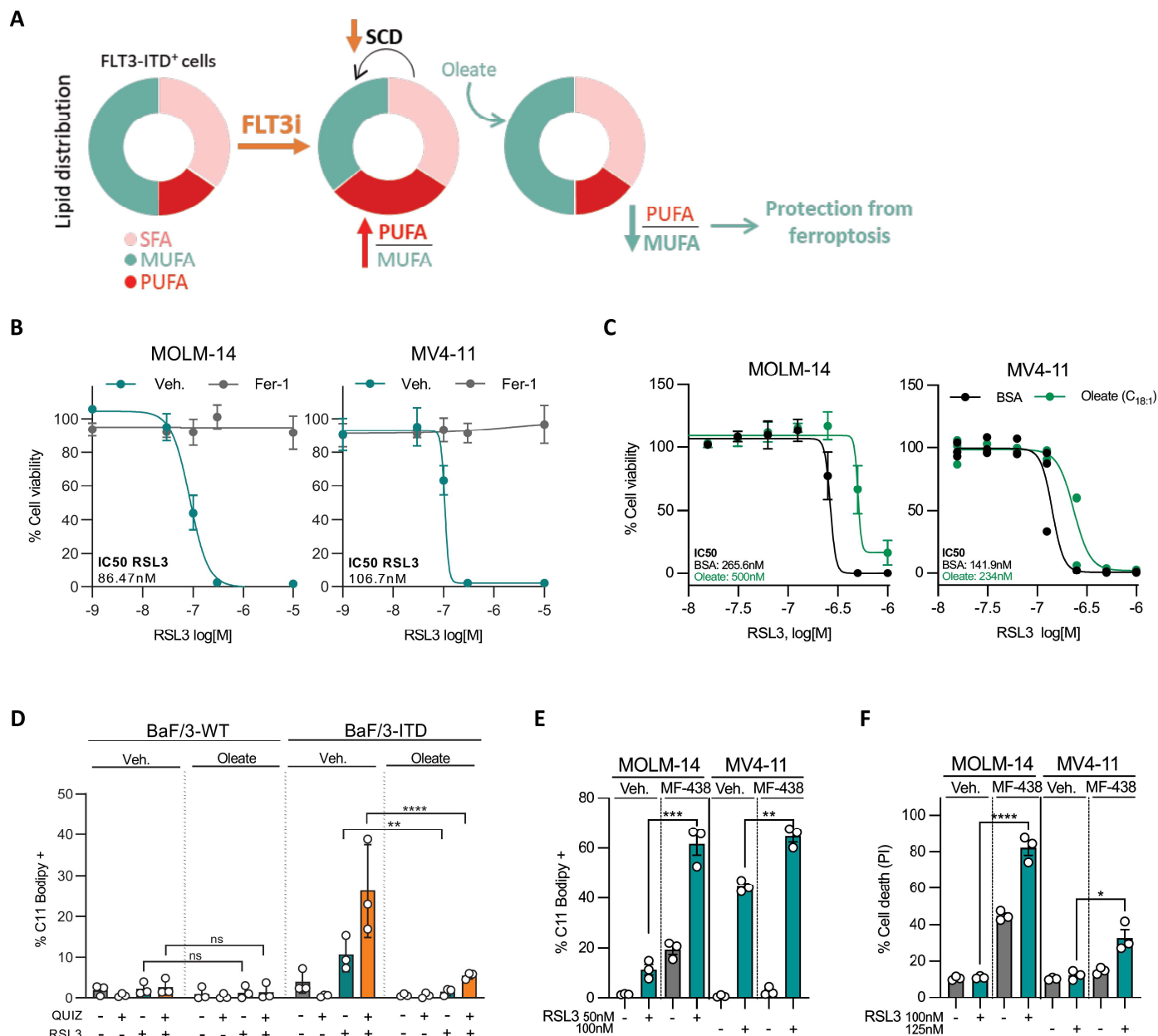


Figure S19

Supplementary Figure S19. Ferroptotic cell death induced by FLT3i is mediated by inhibition of SCD-dependent mono-unsaturated FA synthesis. **A**, Decreased amount of MUFA after TKI treatment related to SCD inhibition is rescued by C_{18:1} oleate supply, protecting AML cells from ferroptotic cell death. **B**, MOLM-14 (*left panel*) and MV4-11 (*right panel*) cells were exposed to dose-range RLS3 with or without 10μM Fer-1 for 48h. Cell viability assays using Alamar Blue. **C**, MOLM-14 (*left panel*) and MV4-11 (*right panel*) cells were exposed to dose-range RLS3 with vehicle (BSA) or 90μM oleate for 48h. Cell viability assays using Alamar Blue. **D**, IL3-dependent Ba/F3 murine hematopoietic cells expressing vectors for FLT3 (FLT3^{WT}) or FLT3-ITD-mutated (FLT3^{ITD}) genes were cultured with vehicle (BSA) or 90μM oleate and treated with or without 1uM RSL3 or 3nM QUIZ alone or in association for 48h. Lipid peroxidation was measured by C11-Bodipy after 48h-treatment. **E-F**, MOLM-14 and MV4-11 cells were incubated with vehicle or 1μM MF-438, combined with vehicle or 50-125nM RSL3. Lipid peroxidation was measured by C11-Bodipy after 12h-treatment (**E**) and cell viability was assessed by PI staining after 24h-treatment (**F**). Vertical bars indicate standard deviations. *: p<0.05; **: p<0.01; ***: p<0.001.

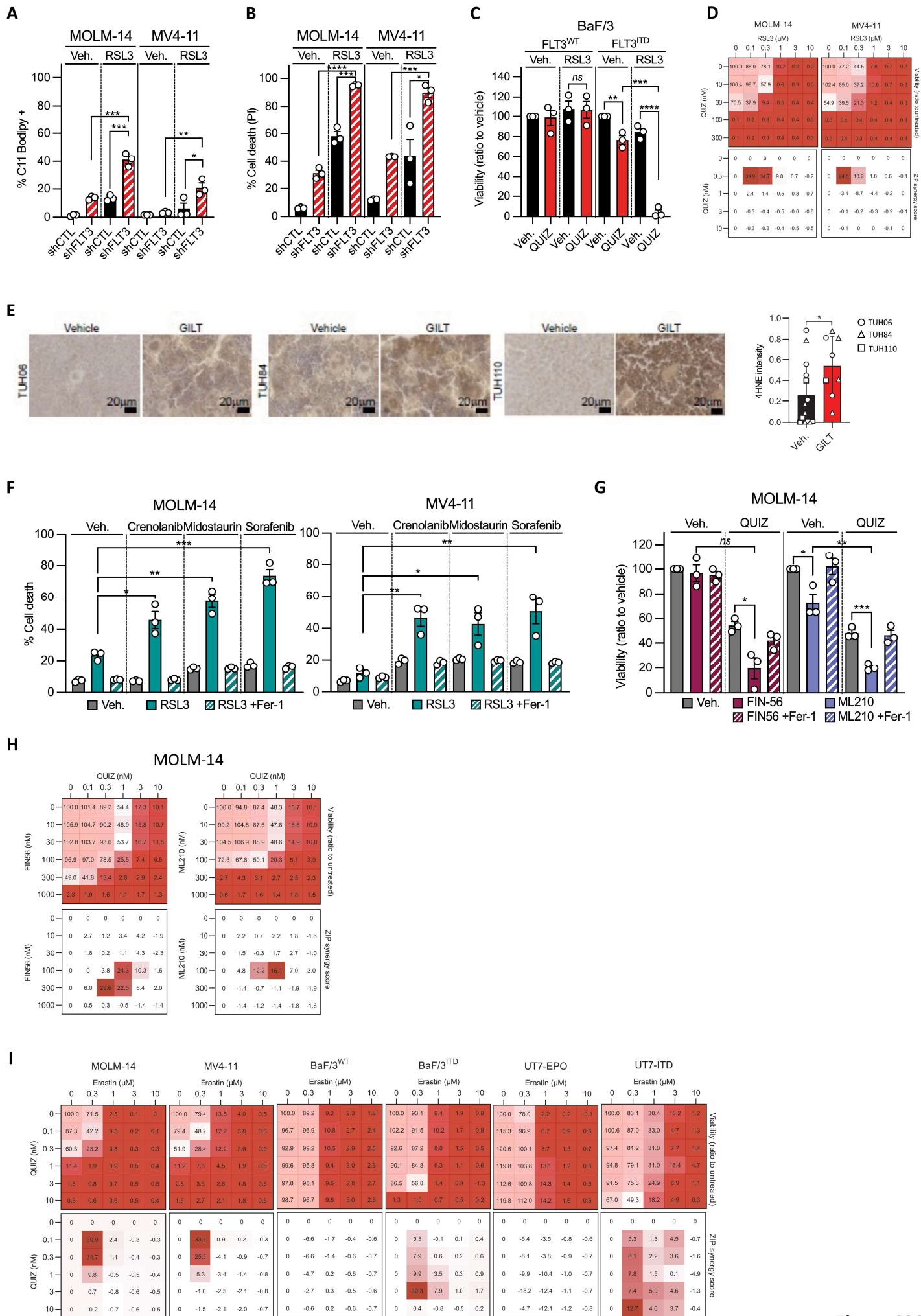


Figure S20

Supplementary Figure S20. FLT3 inhibitors unmask a vulnerability of FLT3-mutant leukemic cells to ferroptosis. **A-B**, MOLM-14 and MV4-11 cells were transduced with CTL or FLT3-targeting shRNAs (dox-inducible, 24h), and incubated with vehicle or 100nM (**A**) or 50nM (**B**) RSL3. **A**, Lipid peroxidation measured by C11-Bodipy staining after 14h. **B**, Cell viability assessed by propidium iodine (PI) staining after 24h. **C**, IL3-dependent Ba/F3 murine hematopoietic cells were transduced with expression vectors for *FLT3* (FLT3^{WT}) or *FLT3*-ITD-mutated (FLT3^{ITD}) genes. FLT3^{WT} and FLT3^{ITD} Ba/F3 cells were grown with and without IL3, respectively. Cells were cultured 48h with vehicle or 3nM QUIZ, and vehicle or 1μM RSL3, and cell viability was assessed using a luminescence-based assay (Alamar Blue). Results are expressed as a ratio to the vehicle conditions. **D**, MOLM-14 and MV4-11 cell lines were incubated with crossed dose-range QUIZ and RSL3 for 48h. Cell viability was measured using a luminescence-based assay (Alamar Blue) and results are expressed as a ratio to the untreated condition for each wheel in a viability matrix (top panel) and using synergy maps using zero interaction potency (ZIP) synergy scores and a threshold ZIP>10 considered as significant (bottom panel). **E**, PDX^{TUH06}, PDH^{TUH84} and PDX^{TUH110} were treated in vivo with vehicle or 30mg/kg/d GILT for 7 days. Next, sternums were harvested and 4-HNE was detected in paraffine-embedded slices using an anti-4HNE antibody. Scales bars (20μM) are provided. **F**, MOLM-14 (*left panel*) and MV4-11(*right panel*) cells were incubated with vehicle, or 10nM and 20nM Crenolanib respectively, or 60nM and 70nM Midostaurin respectively, or 30nM Sorafenib, with vehicle or 100nM RSL3 and with or without 10μM Fer-1 for 24h as indicated. **G**, MOLM-14 cells were incubated with vehicle or 1nM QUIZ, combined without or with 100nM FIN-56 or 100nM ML-210 and with or without 10μM Fer-1 for 48h as indicated, and cell viability was assessed using a luminescence-based assay (Alamar Blue). **H**, Dose-response matrices in MOLM-14 cells incubated with crossed dose-range QUIZ and FIN56 for 48h. Cell viability was measured using a luminescence-based assay (Alamar Blue) and results are expressed as a ratio to the untreated condition for each wheel in a viability matrix (top panel) and using synergy maps using zero interaction potency (ZIP) synergy scores and a threshold ZIP>10 considered as significant (bottom panel). **I**, Dose-response matrices for QUIZ and erastin combination in MOLM-14, MV4-11, Ba/F3 transduced with FLT3 wild-type (WT) or FLT3-ITD alleles (Ba/F3^{WT} and Ba/F3^{ITD}, respectively), parental UT7 (UT7 EPO) and UT7 transduced with FLT3-ITD (UT7-ITD) cell lines. Cell viability was measured after 48h using a luminescence-based assay (Alamar Blue) and results are expressed as a ratio to the untreated condition for each wheel in a viability matrix (top panel) and using synergy maps using zero interaction potency (ZIP) synergy scores and a threshold ZIP>10 considered as significant (bottom panel).

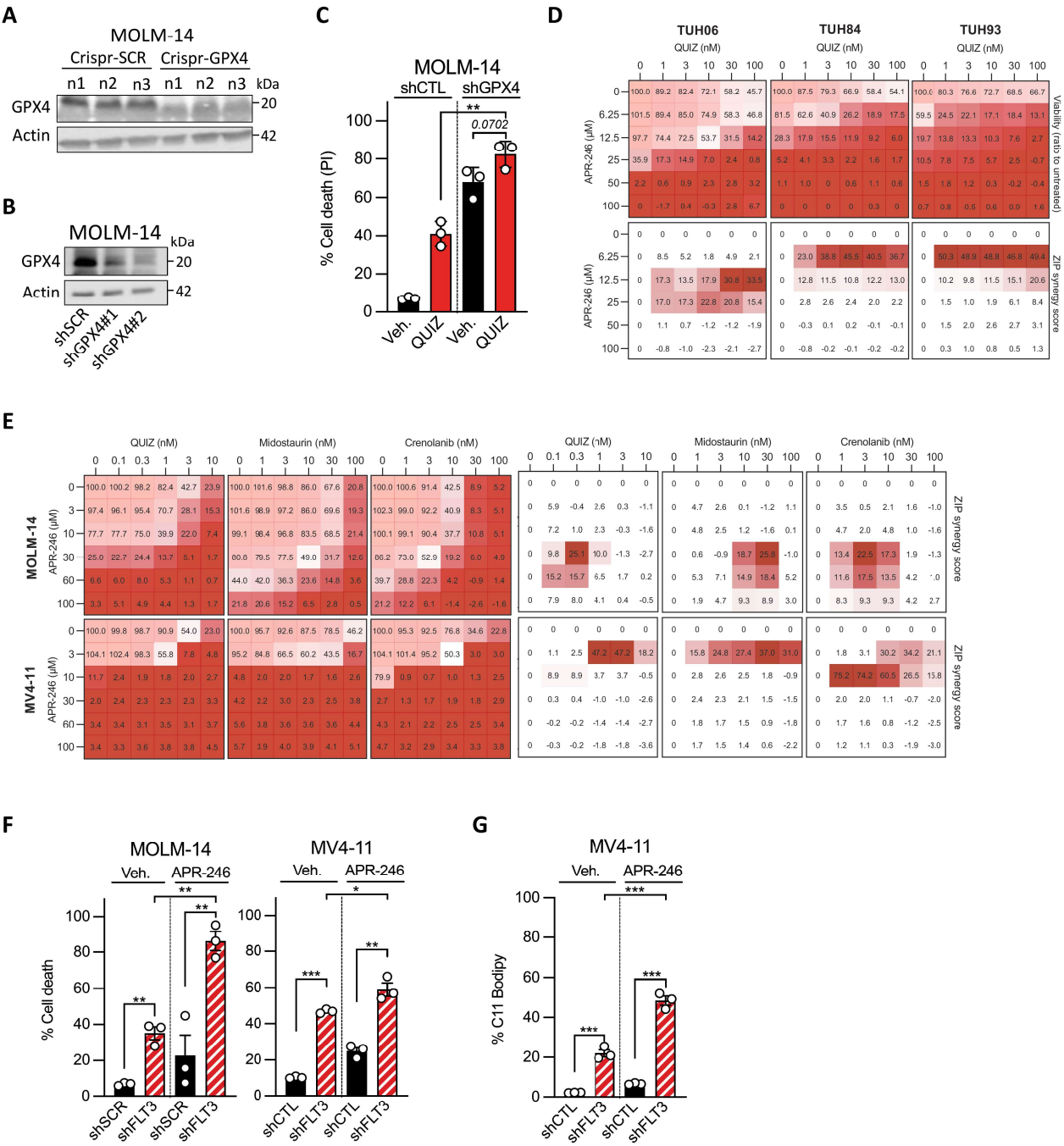


Figure S21

Supplementary Figure S21. Lipid redox stress induction by GPX4 inhibition primed FLT3i activity in FLT3-ITD AML cells. **A**, Representative western blot of MOLM-14 cells transduced with CTL or GPX4-targeting CRISPR-Cas9 vectors, and cultured with 10 μ M Fer-1 to avoid cell death induced by GPX4 ablation. **B-C**, MOLM14 cells were transduced with dox-inducible shCTL or shGPX4 and cultured with dox for 72h. **B**, Total protein extracts were immunoblotted with the indicate antibodies. **C**, Cell death was measured using PI staining in shCTL versus shGPX4 MOLM-14 cells treated with vehicle or 3nM QUIZ for 48h. **D**, Synergy matrices in cells from PDX^{TUH06}, PDH^{TUH84} and PDX^{TUH110} incubated with QUIZ and APR-246 for 48h. Cell viability was measured using a luminescence-based assay (Alamar Blue) and results are expressed as a ratio to the untreated condition for each wheel in a viability matrix (*left panels*) and using synergy maps using zero interaction potency (ZIP) synergy scores and a threshold ZIP>10 considered as significant (*right panels*). **E**, Synergy matrices in MOLM-14 and MV4-11 cells incubated with QUIZ, Midostaurin or Crenolanib and APR-246 for 48h. Cell viability was measured using a luminescence-based assay (Alamar Blue) and results are expressed as a ratio to the untreated condition for each wheel in a viability matrix (*left panels*) and using synergy maps using zero interaction potency (ZIP) synergy scores and a threshold ZIP>10 considered as significant (*right panels*). **F-G**, MOLM-14 and MV4-11 cells transduced with CTL or FLT3-targeting shRNAs were cultured with vehicle or 30-5 μ M APR-246, respectively, for 24h. **F**, Cell viability measured using PI staining. **G**, Lipid peroxidation measured using C11 Bodipy staining. Vertical bars indicate standard deviations. **p<0.01, ***p<0.001.

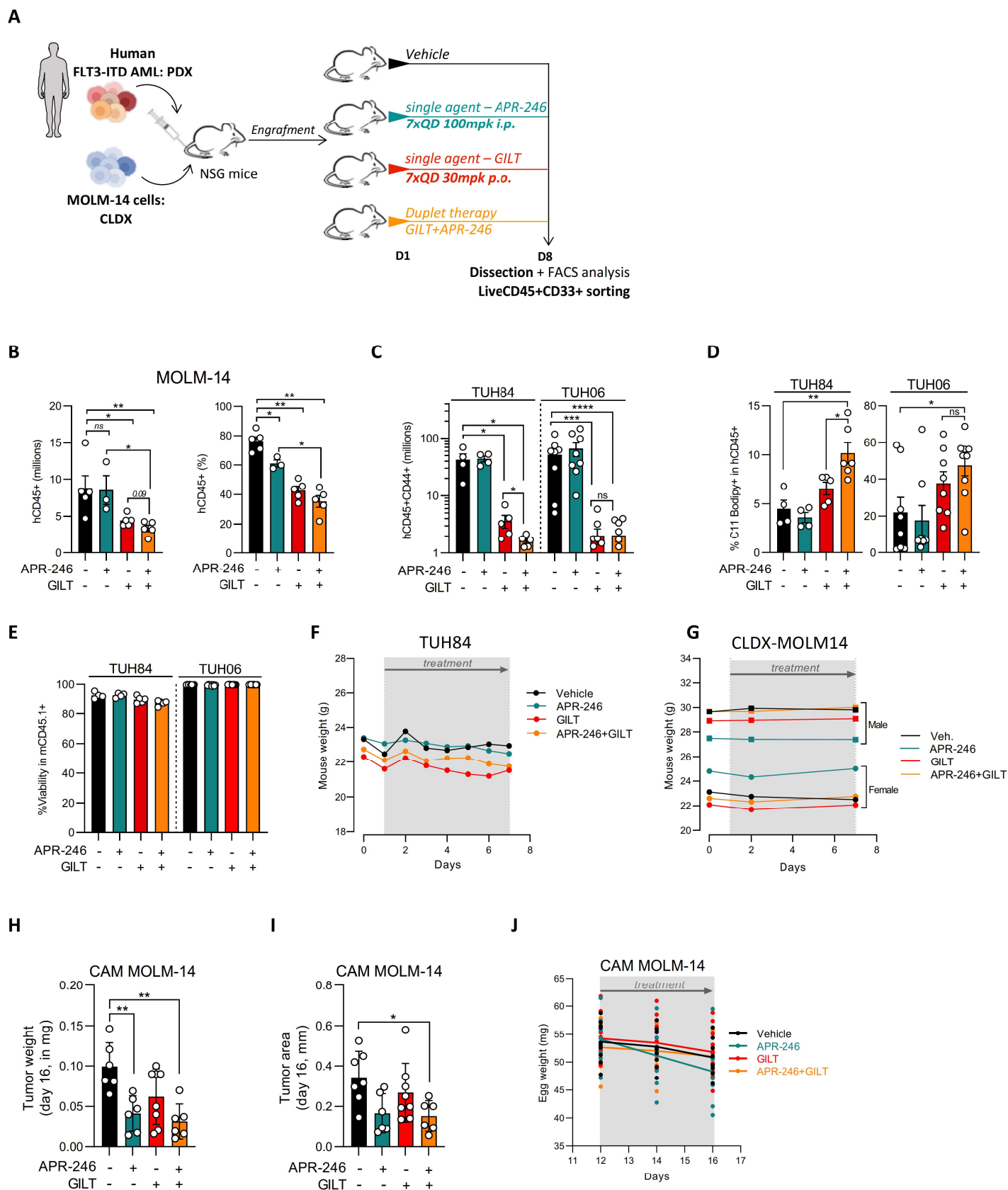


Figure S22

Supplementary Figure S22. Combined treatment with GILT and APR-246 in preclinical AML models *in vivo*. **A**, PDX and cell line-derived xenograft (CLDX) assays using MOLM-14 cell line. After engraftment, mice were treated with vehicles, 100mg/kg/day APR-246 given by i.p., 30mg/kg/day GILT for PDX and 10mg/kg/day GILT for CLDX given by oral gavage, or combination of these two agents for 7 days (n= 5 to 8 mice per group). **B**, Number of viable hCD45+ cells (*left panel*) and percentage of viable hCD45+ AML cells (*right panel*) in bone marrow of CLDX MOLM14 mice. **C**, Number of viable hCD45+CD44+ AML cells in bone marrow and spleen from PDX^{TUH84} and PDX^{TUH06}. **D**, Lipid peroxidation measured by C11-bodipy staining in hCD45+ AML cells from PDX^{TUH84} and PDX^{TUH06}. **E**, Viability measurement of murine hematopoietic mCD45.1+ cells from PDX^{TUH84} (*left panel*) and PDX^{TUH06} (*right panel*) using Annexin V staining. **F**, Mice weight monitoring during treatment of PDX^{TUH84}. **G**, Mice weight monitoring during MOLM-14 CLDX assays. Weight is normalized to the initial weight by mouse before treatment start. **H-J**, Chick chorioallantoic membrane (CAM) assay of MOLM-14 transplantation treated with vehicle, 30μM APR-246, 30nM GILT or combination for 7 days. Tumor weight (**H**) and area (**I**) in eggs collected at day 16. **J**, Eggs weight monitoring during treatment.