Tumor cell-derived microparticles induced by methotrexate augment T-cell antitumor responses by down-regulating expression of PD-1 in neutrophils

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Supplementary Figures S1–S7 and supplementary table S1



Supplementary Fig S1 related to Fig 1.

(A-B) MTX-MPs were isolated from H22 tumor cells, size distribution was measured by Malvern Zetasizer Nano ZS90 particle size analyzer (A), morphology was analyzed by scanning electron microscope (B), and annexin V (C) was analyzed by flow cytometry. (D) 5×10^7 H22 cells derived MTX-MPs were prepared and the drugs include were analyzed by HPLC. Drugs were quantified by using Ultimate 3000 HPLC system in a C18 column, and drugs concentration were calculated according to standard curve. (E) The stability of MTX-MPs was analyzed under different conditions including pH5.5, pH9.0, explored to sunlight and room temperature (RT) for a week, and SDS or proteinase K for 30 min. MTX-MPs were then recollected and mixed with 3µm beads for flow cytometric analysis, P1 represent beads and P2 represent MTX-MPs. (F) Sterility of MTX-MPs suspension were analyzed. MTX-MPs suspension were added to Trypticase Soy Broth (TSB) for bacteria growth, and E. coli as a positive control and Water For Injection (WFI) as negative control. For all graphs, data present mean \pm SEM, *****P*<0.0001, ns, not statistically significant by One way ANOVA (E).



Supplementary Fig S2 related to Fig 1

 3×10^5 H22 tumor cells were injected subcutaneously, when tumor size reach 5×5 mm², the mice were i.v injected with MTX-MPs daily for 3 times, the neutrophils in tumor tissue were analyzed using immunofluorescence staining. Scale bar 20 μ m.



Supplementary Fig S3 related to Fig 1

(A - B) H22 derived MTX-MPs were incubated with CFSE-labeled CD8⁺ T cells for 3 days, the proliferation (A) and IFN- γ releasing (B) were determined by flow cytometry. For all graphs, data present mean \pm SEM, ns, not statistically significant by unpaired Student's t-test (A and B).



Supplementary Fig S4 related to Fig 2

(A) Neutrophils were isolated from H22 ascites mice, and incubated with MTX-MPs for 6 hours, the expression of PD-L1 was analyzed by flow cytometry. (B) Same as (A),

the neutrophils were pretreated with amiloride hydrochloride (Ami, 100 μ M) for 1 hour and then incubated with PKH26-labeled MTX-MPs for 6 hours, the uptake of MTX-MPs was analyzed by flow cytometry. **(C-D)** Neutrophils were isolated from H22 ascites mice, and incubated with MTX-MPs in the presence of Ami **(C)** or DUB-IN-1 (DUB, 1 μ M) **(D)** for 6 hours, the expression of PD-1 was analyzed by flow cytometry. **(E-F)** Bone-marrow neutrophils were isolated from H22 tumor-bearing mice, and incubated with MTX-MPs **(E)** or NH4Cl (1mM) **(F)**, the pH changed was analyzed using lysosensor green. **(G)** Neutrophils were isolated from H22 ascites mice, and incubated with MTX-MPs in the presence of NH4Cl for 6 hours, the expression of PD-1 was analyzed by flow cytometry. For all graphs, data present mean \pm SEM, ****P*<0.001, *****P*<0.0001, ns, not statistically significant by Student's t-test (A, E and F) or One way ANOVA (C, D and G).



Supplementary Fig S5 related to Fig 3

(A-B) Neutrophils were isolated from bone marrow of healthy mice and cultured with CM of CD3/CD28 antibody-activated CD8⁺ T cells in the presence of PD-L1 antibody (A) or IFN- γ (20 ng/mL), IL-2 (20 ng/mL) (B) for 12 hours, the expression of PD-1 in neutrophils was analyzed by flow cytometry. (C-D) Neutrophils were isolated from bone marrow of healthy mice and cultured with lactic acid for 12 hours, the expression of PD-L1(C) were determined by flow cytometry, or cultured with LLC CM, or LLC CM plus CHC for 12 hours, the expression of PD-1 (D) was determined by flow cytometry. For all graphs, data present mean \pm SEM, *****P*<0.0001, ns, not statistically significant by Student's t test (C) or One way ANOVA (A, B and D).



Supplementary Fig S6 related to Fig 5

(A-B) 1×10^5 H22 tumor cells were i.p injected to BALB/c mice (A), or 1×10^6 LLCs were i.v injected to C57BL/6 mice (B) for 10 days, the expression of PD-1 in neutrophil in spleen and bone marrow were determined by flow cytometry. For all graphs, data present mean \pm SEM, ***P*<0.01, **** *P*<0.0001, by unpaired Student's t-test (A and B).



Supplementary Fig S7 related to Fig 6

(A-C) 1×10^{6} LLCs were i.v injected to C57BL/6 mice for 7 days, then PBS, MTX-MPs, anti-PD-L1 (20 µg/g, daily for 4days) or MTX-MPs (daily for 5 days) combined with anti-PD-L1 antibody were i.v injected, IL-2 (A), CD25 (B) and CD69 (C) expressed in tumor-infiltrating CD8⁺ T cells were analyzed by flow cytometry. *****P*<0.0001, ns, not statistically significant by One way ANOVA.

Supplementary Table S1 Primer sequences

| Gene | Primer | Primer sequence |
|-------------|--------|------------------------|
| Mouse Arg1 | FW | CAAGACAGGGCTCCTTTCAG |
| | RV | TGGCTTATGGTTACCCTCC |
| Mouse Nos2 | FW | GCAGCTTGTCCAGGGATTCT |
| | RV | GCAGCTTGTCCAGGGATTCT |
| Mouse Actb | FW | GGCTGTATTCCCCT-CCATCG |
| | RV | CCAGTTGGTAACAATGCCATGT |
| Mouse Il10 | FW | TGGACTCCAGGACCTAGACA |
| | RV | CGGAGAGAGGTACAAACG |
| Mouse Elane | FW | CATGGCCCTTGGCAGACTAT |
| | RV | AGTTCCTGGCAATGAGGGTG |