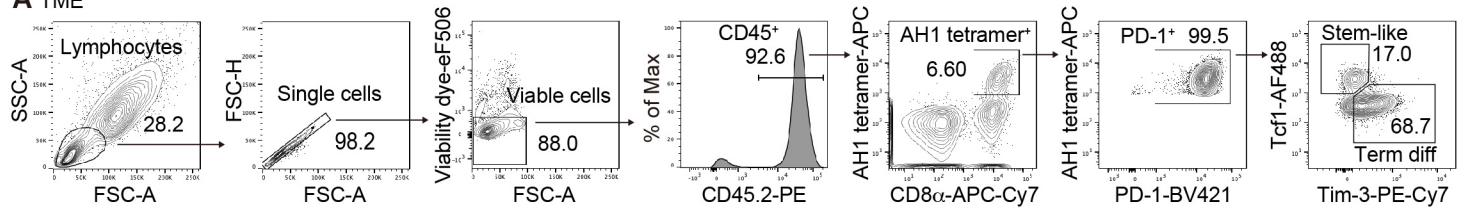
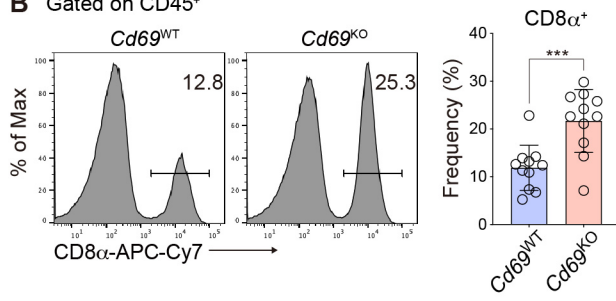


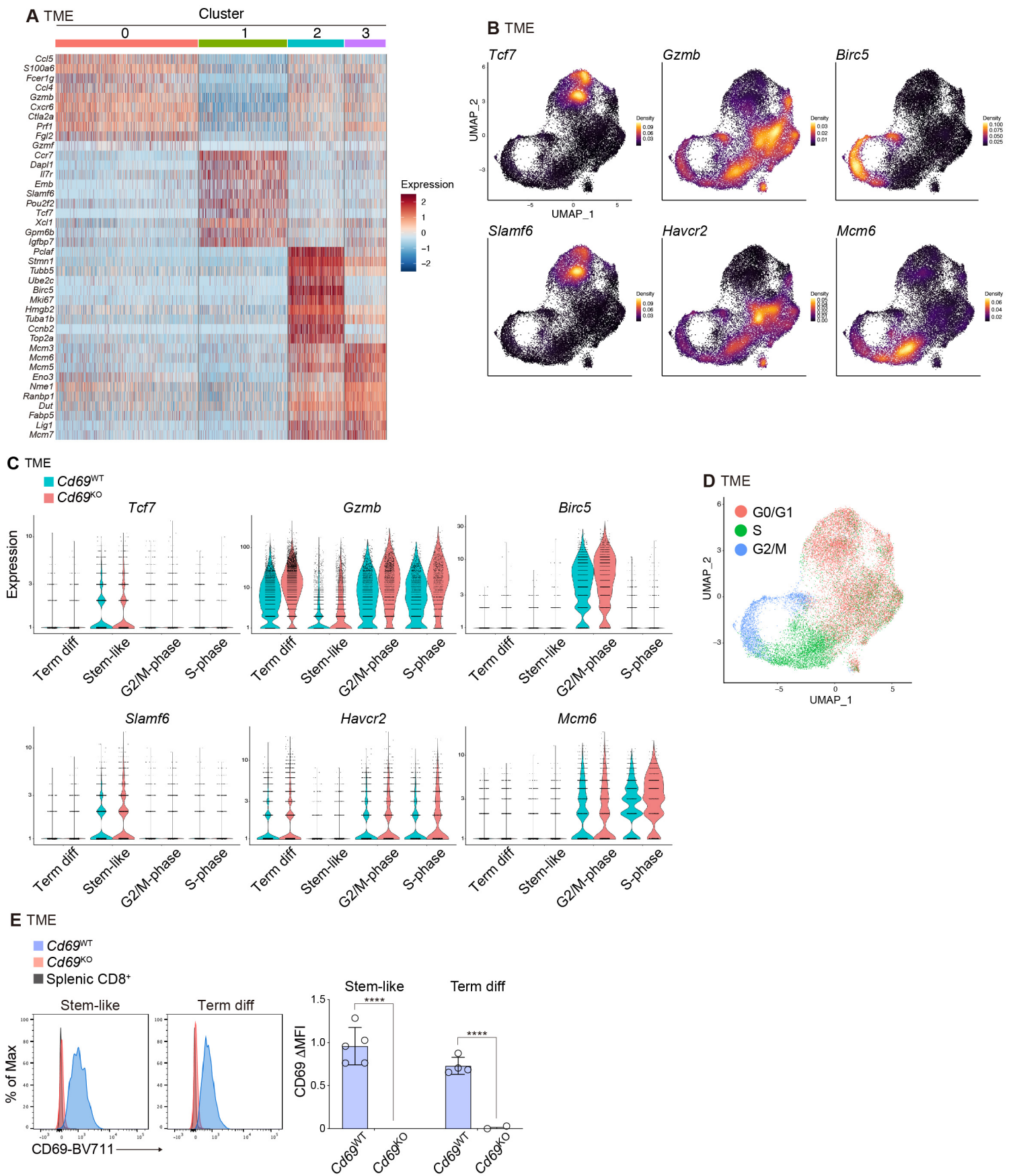
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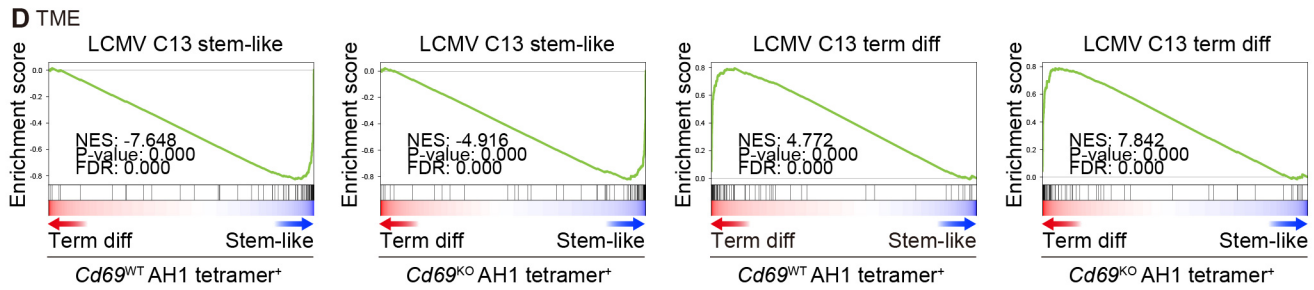
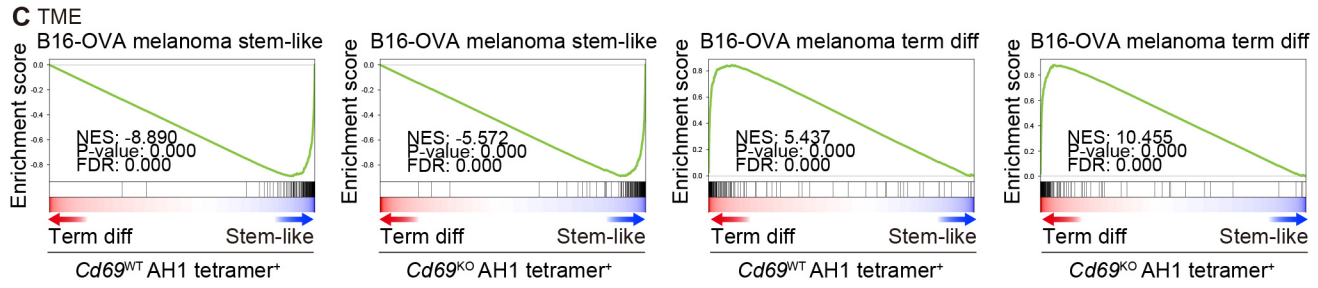
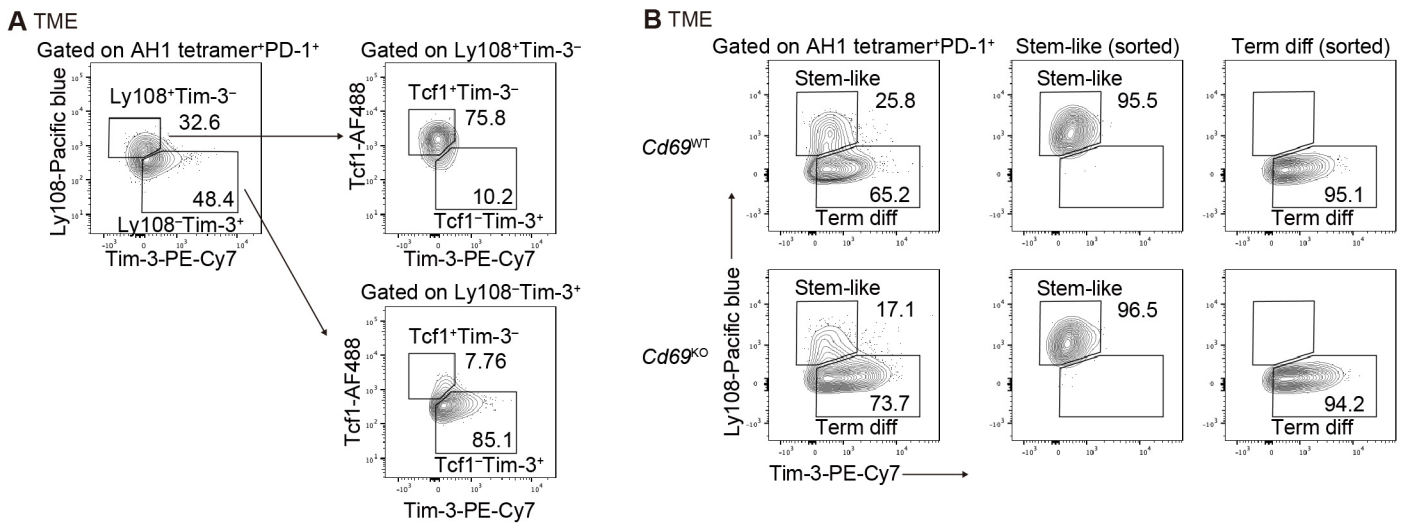
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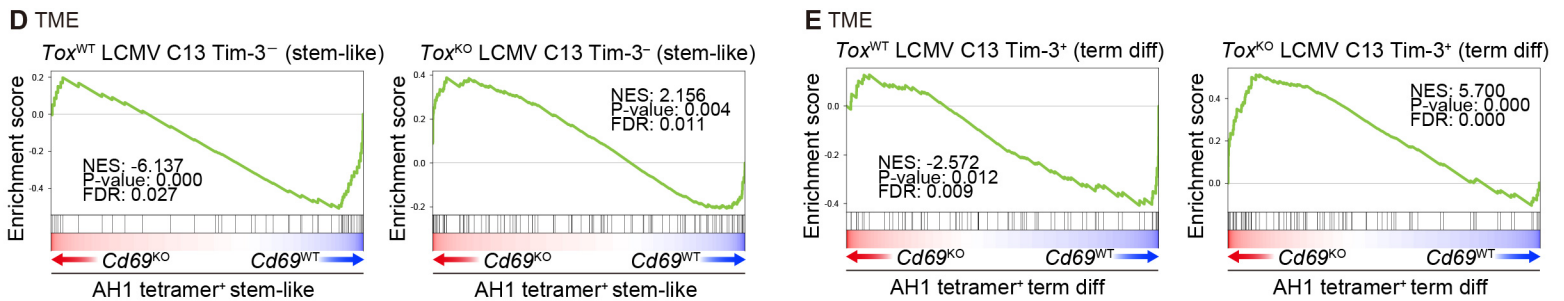
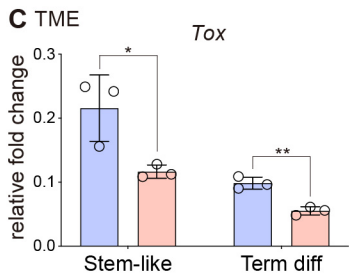
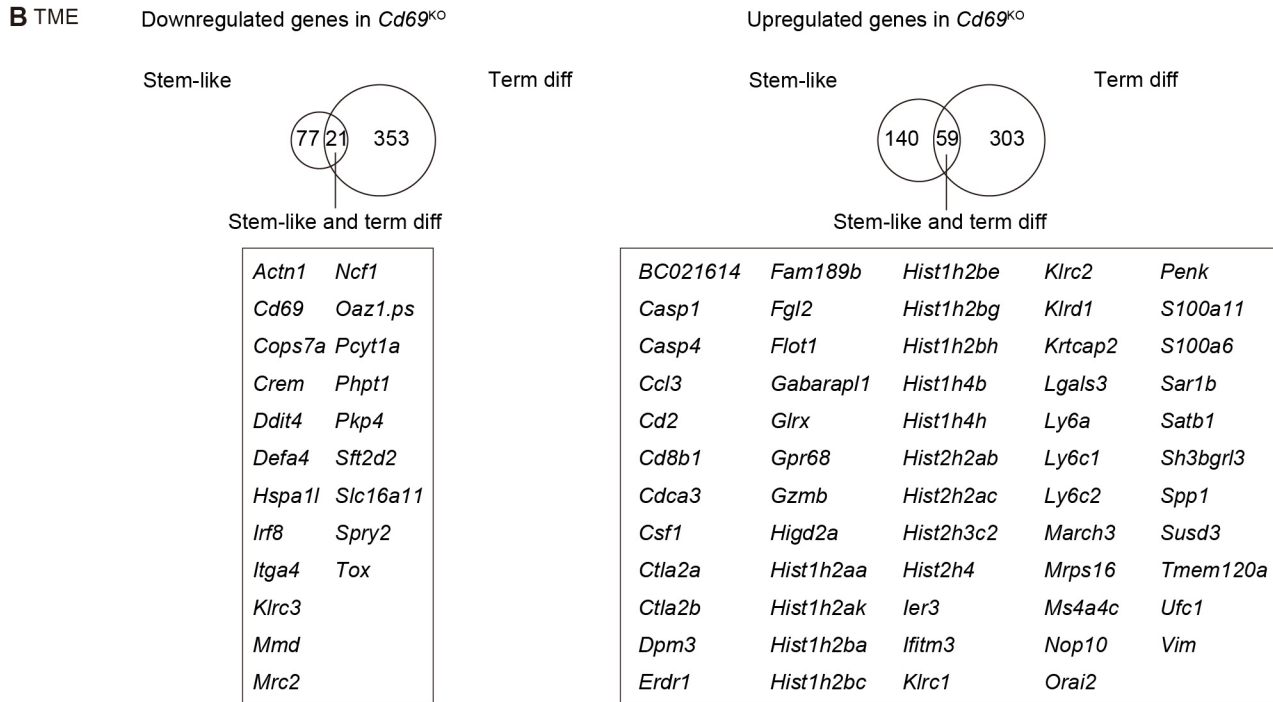
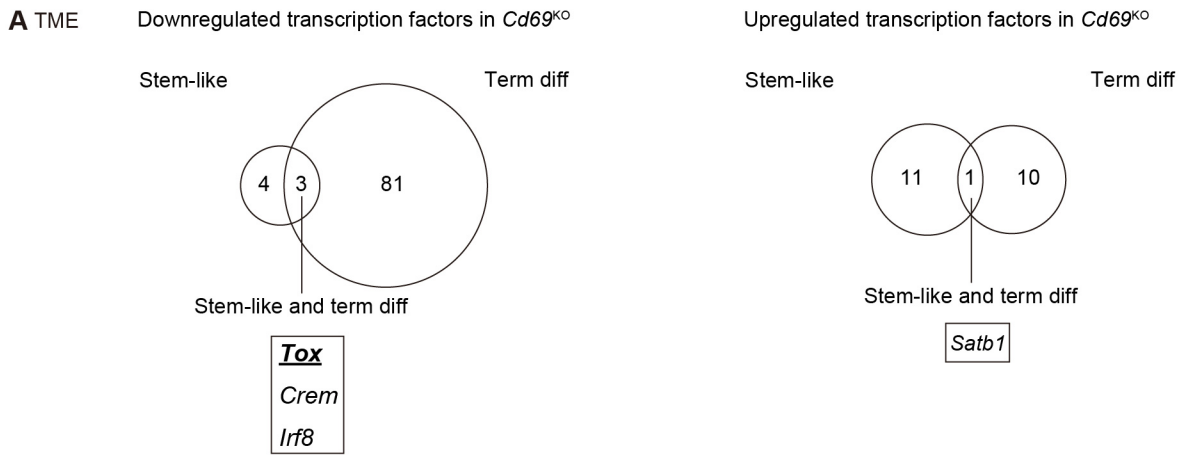
Supplementary Figure S1. Flow cytometry of intratumoral tumor-specific CD8⁺ T cells. **A**, A representative flow cytometry gating strategy for intratumoral CD8⁺ T cell subpopulations. CT26 cells were subcutaneously inoculated into 7-week-old female BALB/c mice. Intratumoral AH1 tetramer⁺ CD8⁺ T cells were analyzed at 14 days after inoculation. **B**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 7-week-old female BALB/c mice. Intratumoral CD8⁺ T cells were analyzed at 14 days after CT26 cell inoculation. At least n=6 per group. The number indicates the percentage of gated cells. The frequency of CD8⁺ T cells in CD45⁺ cells is shown in representative flow plots (left) and a summary of the experiments (right). An unpaired two-sided Student's *t*-test or Welch's *t*-test was used for the analysis of the flow cytometry data. *** P<0.001. Data are pooled from two experiments with at least five mice (**B**).



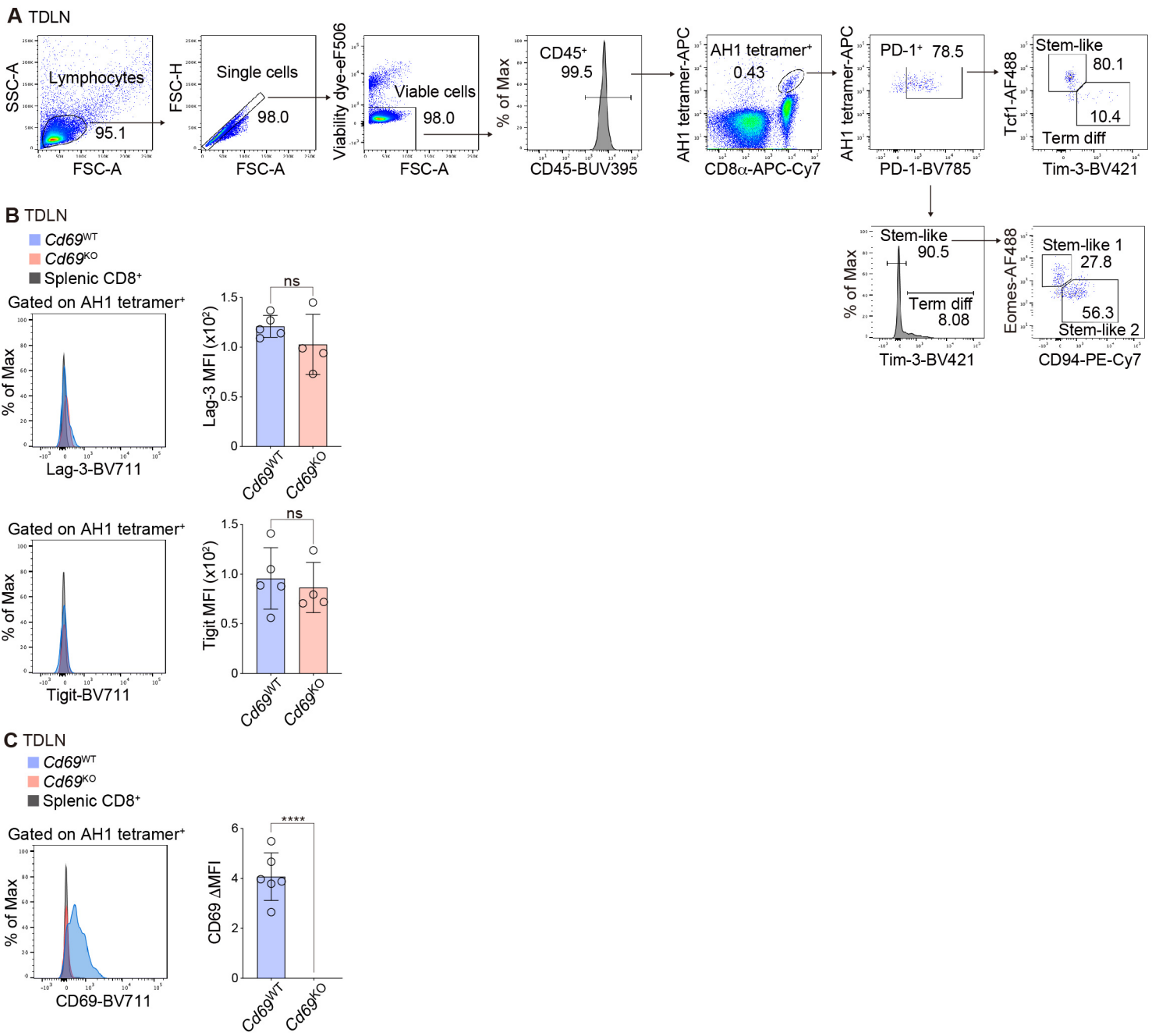
Supplementary Figure S2. scRNA-seq of intratumoral tumor-specific CD8⁺ T cells. **A**, scRNA-seq. CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 7-week-old female BALB/c mice. Intratumoral AH1 tetramer⁺ CD8⁺ T cells were isolated at 14 days after inoculation. Experiments were performed in duplicate. Pool of four mice in 1 group. A UMAP projection of *Cd69*^{WT} (n=9,870 and 4,994 cells) and *Cd69*^{KO} (n=6,090 and 7,551 cells) cells that were assigned to 4 clusters and color-coded based on the clusters, determined by Seurat. A heatmap of the top 10 genes expressed in each cluster as defined in Fig. 1F. The columns correspond to the cells, the rows correspond to the genes. Cells are grouped by clusters. The color scale is based on a z-score distribution from -2 (blue) to 2 (red). **B**, Single-cell transcript levels of the indicated genes illustrated in the UMAP plots. **C**, Violin plots showing the expression of the indicated genes in cells from each cluster as defined in Fig. 1F. The violin represents the probability density at each value. Each dot represents a single cell. **D**, The cell-cycle phase (G0/G1, S, or G2/M) of each cell was determined by Seurat and is shown in the UMAP plots. **E**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 7-week-old female BALB/c mice. Intratumoral AH1 tetramer⁺ CD8⁺ T cells were analyzed at 14 days after inoculation. At least n=4 per group. Delta MFI of CD69 (delta MFI was calculated by subtracting splenic CD8⁺ MFI from AH1-specific CD8⁺ T MFI in CD8⁺ T cell subpopulations). A representative histogram (left) and summary of the experiments (right). An unpaired two-sided Student's *t*-test or Welch's *t*-test was used for the analysis of the flow cytometry data. **** P<0.0001. ns: not significant. Representative data from three experiments with at least four mice are shown (**E**).



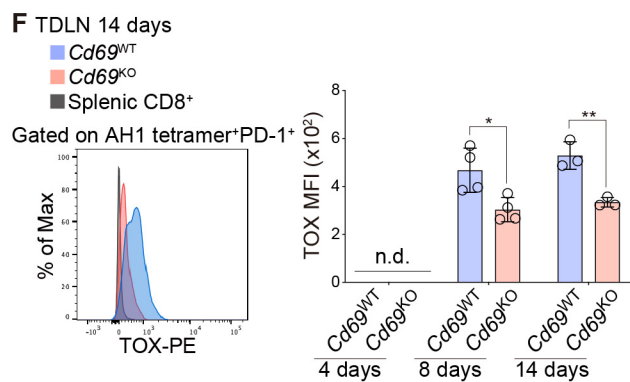
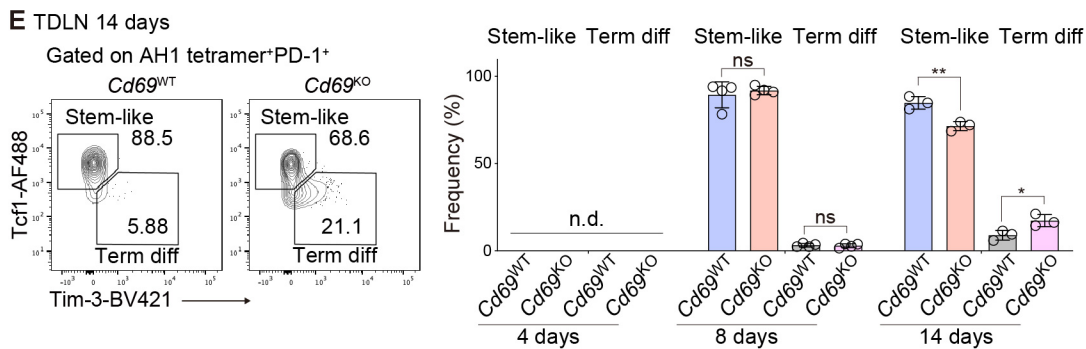
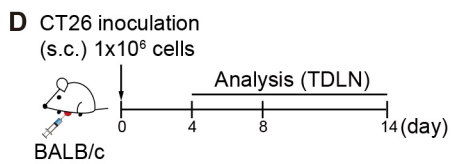
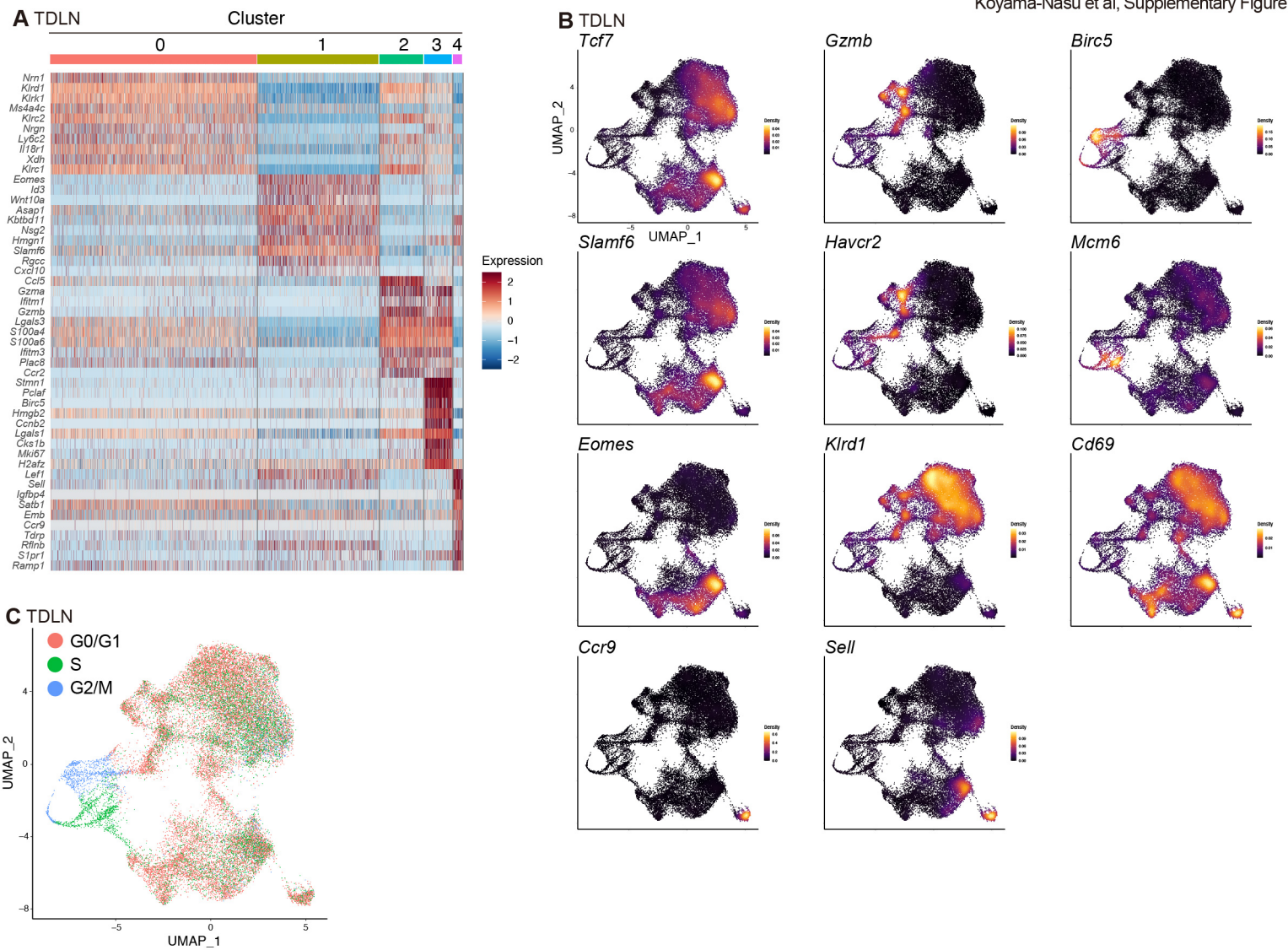
Supplementary Figure S3. Bulk RNA-seq of intratumoral tumor-specific CD8⁺ T cells. **A**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 7-week-old female BALB/c mice. Fourteen days after inoculation, the intratumoral AH1 tetramer⁺ CD8⁺ T cell subpopulations were analyzed. For live cell sorting, the cell surface marker Ly108 was used instead of Tcf1 (1). **B**, The intratumoral AH1 tetramer⁺ CD8⁺ T cell subpopulations were sorted and subjected to RNA-seq. Experiments were performed in duplicate. Pool of four mice in 1 group. A representative flow cytometry gating strategy for Ly108⁺ stem-like and Tim-3⁺ terminally differentiated CD8⁺ T cells is shown. The number indicates the percentage of gated cells. **C**, A GSEA of the transcriptional signature from stem-like or terminally differentiated CD8⁺ T cells in B16-OVA melanoma in the ranked list of genes differentially expressed by AH1 tetramer⁺ terminally differentiated CD8⁺ T cells versus stem-like CD8⁺ T cells in *Cd69*^{WT} and *Cd69*^{KO} mice. **D**, A GSEA of the transcriptional signature from stem-like or terminally differentiated CD8⁺ T cells in LCMV clone-13 infection in the ranked list of genes differentially expressed by AH1 tetramer⁺ terminally differentiated CD8⁺ T cells versus stem-like CD8⁺ T cells in *Cd69*^{WT} and *Cd69*^{KO} mice. The gene expression profiles of CD8⁺ T cell subpopulations in B16-OVA melanoma and LCMV clone-13 infection were obtained from Miller et al. (1).



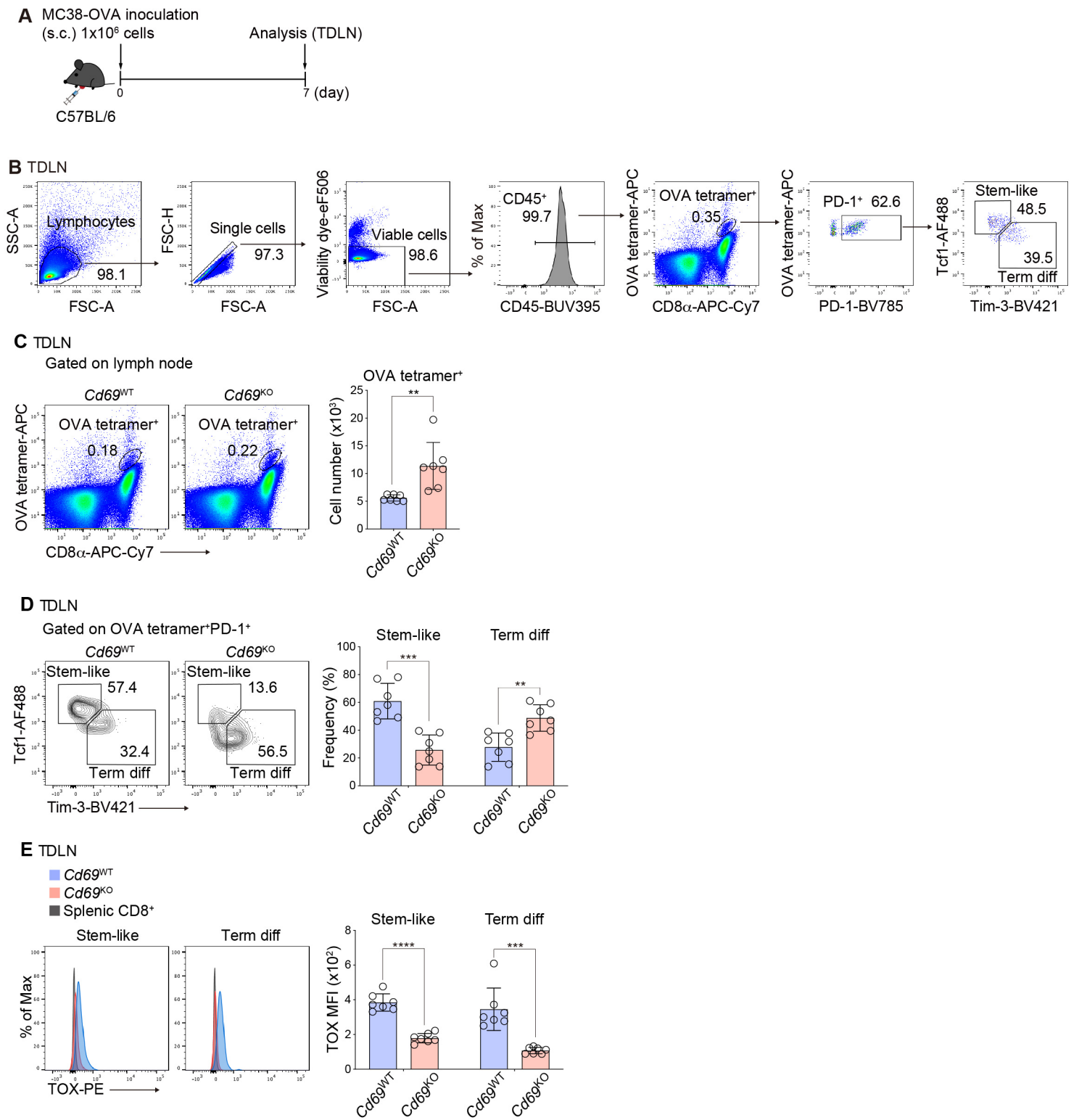
Supplementary Figure S4. CD69 controls the expression of the transcription factor TOX and TOX-mediated transcriptional program. **A**, A Venn diagram showing the overlap among genes encoding transcription factors downregulated in *Cd69*^{KO} stem-like CD8⁺ T cells and those downregulated in *Cd69*^{KO} terminally differentiated CD8⁺ T cells (left), showing the overlap among genes encoding transcription factors upregulated in *Cd69*^{KO} stem-like CD8⁺ T cells and those upregulated in *Cd69*^{KO} terminally differentiated CD8⁺ T cells (right). **B**, A Venn diagram showing the overlap among genes downregulated in *Cd69*^{KO} stem-like CD8⁺ T cells and those downregulated in *Cd69*^{KO} terminally differentiated CD8⁺ T cells (left), showing the overlap among genes upregulated in *Cd69*^{KO} stem-like CD8⁺ T cells and those upregulated in *Cd69*^{KO} terminally differentiated CD8⁺ T cells (right). **C**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 7-week-old female BALB/c mice. Intratumoral AH1 tetramer⁺ CD8⁺ T cell subpopulations were sorted at 14 days after CT26 cell inoculation and subjected to Quantitative RT-PCR. n=3 per group. The graph represents relative fold change of *Tox* mRNA. **D**, A GSEA of the transcriptional signature from *Tox*^{WT} (left) or *Tox*^{KO} (right) Tim-3⁻ (stem-like) cells in LCMV clone-13 infection in the ranked list of genes differentially expressed in AH1 tetramer⁺ stem-like CD8⁺ T cells in *Cd69*^{KO} mice versus *Cd69*^{WT} mice. **E**, A GSEA of the transcriptional signature from *Tox*^{WT} (left) or *Tox*^{KO} (right) Tim-3⁺ (terminally differentiated) cells in LCMV clone-13 infection in the ranked list of genes differentially expressed in AH1 tetramer⁺ terminally differentiated CD8⁺ T cells in *Cd69*^{KO} mice versus *Cd69*^{WT} mice. The gene expression profiles of *Tox*^{WT} and *Tox*^{KO} CD8⁺ T cell subpopulations in LCMV clone-13 infection were obtained from Alfei et al. (30). NES, normalized enrichment score. FDR, false discovery rate. An unpaired two-sided Student's *t*-test was used for the analysis of the quantitative RT-PCR data. * P<0.05, ** P<0.01.



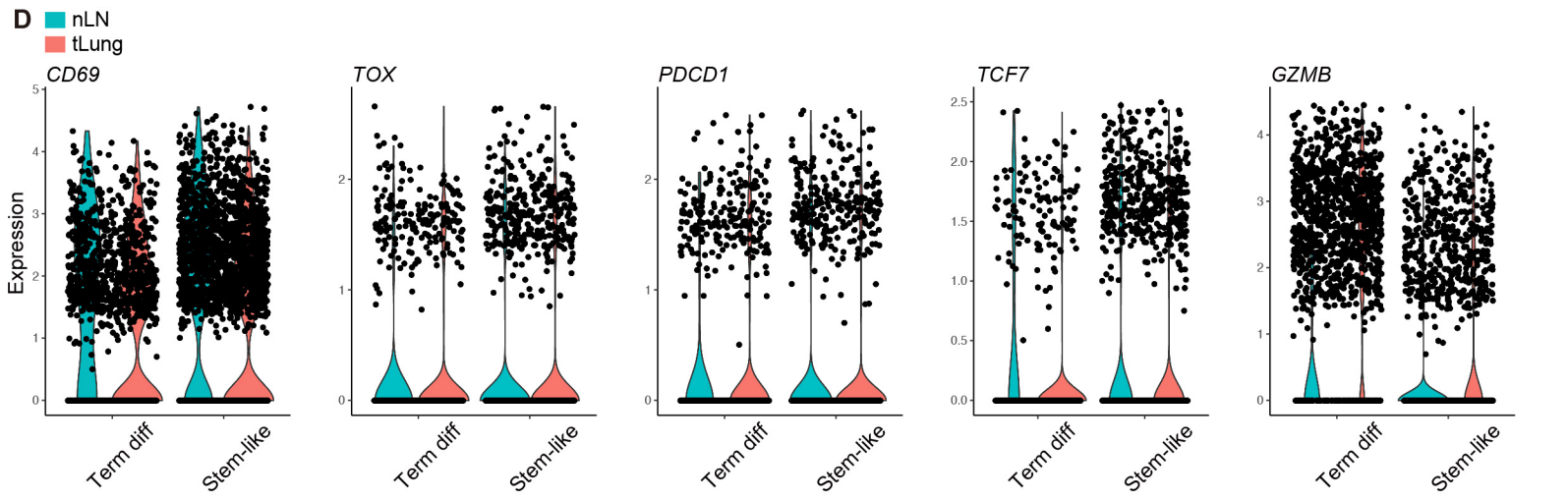
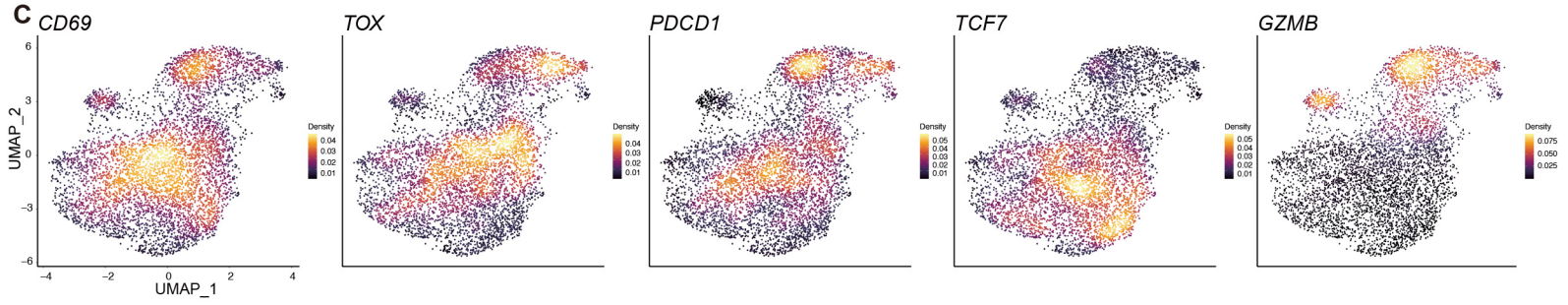
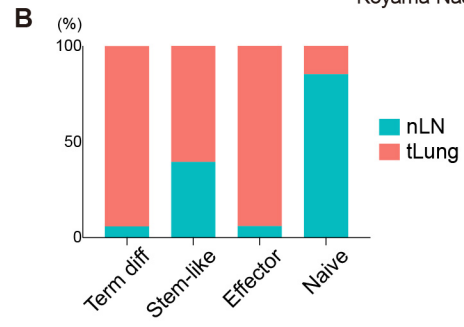
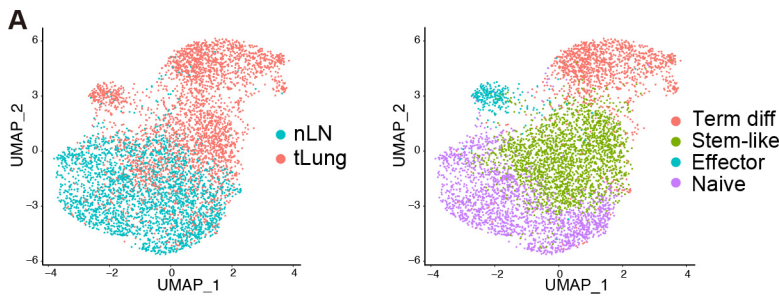
Supplementary Figure S5. Flow cytometry of tumor-specific CD8⁺ T cells in TDLNs. **A**, Representative flow cytometry gating strategy for CD8⁺ T cell subpopulations in TDLNs. CT26 cells were subcutaneously inoculated into 8-week-old female BALB/c mice. AH1 tetramer⁺ CD8⁺ T cells in TDLNs were analyzed at 14 days after inoculation. **B**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 8-week-old female BALB/c mice. AH1 tetramer⁺ CD8⁺ T cells in TDLNs were analyzed at 14 days after inoculation. At least n=4 per group. MFI of the indicated proteins in CD8⁺ T cells. A representative histogram (left) and summary of the experiments (right). Splenic CD8⁺ cells are shown as a control. **C**, Delta MFI of CD69 (delta MFI was calculated by subtracting splenic CD8⁺ MFI from AH1-specific CD8⁺ T MFI). An unpaired two-sided Student's *t*-test was used for the analysis of the flow cytometry data. **** P<0.0001. ns: not significant. Representative data from two experiments with at least four mice are shown (**B-C**).



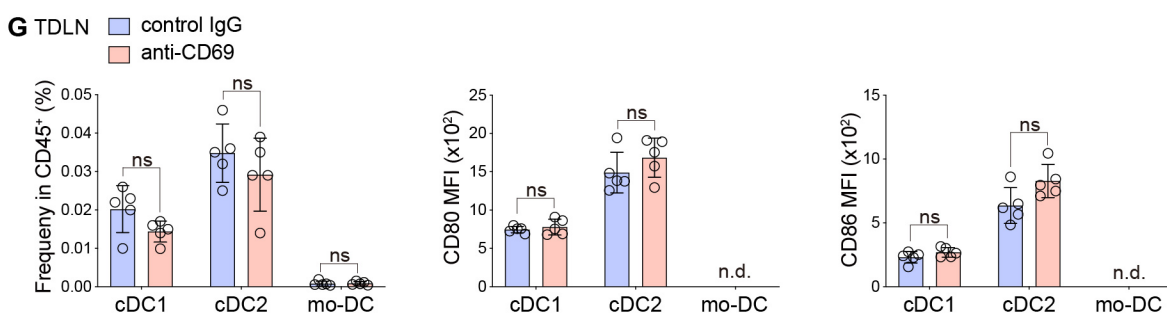
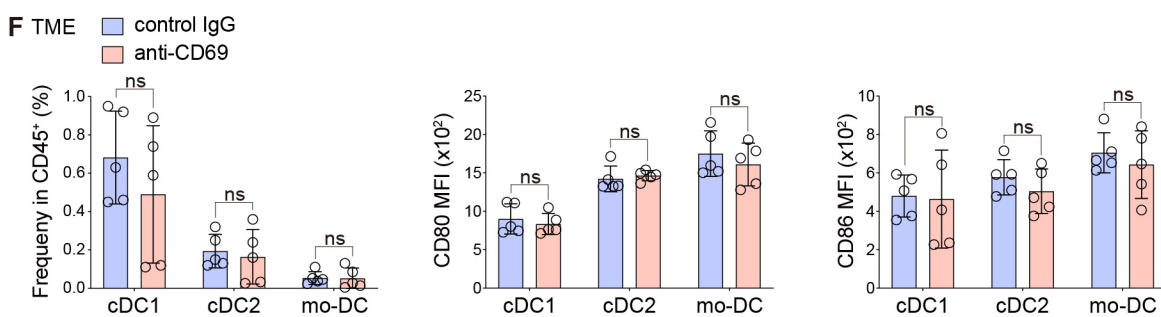
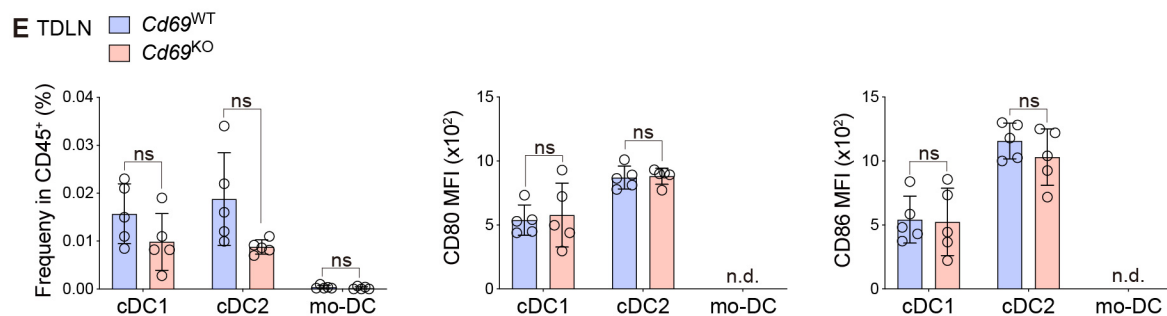
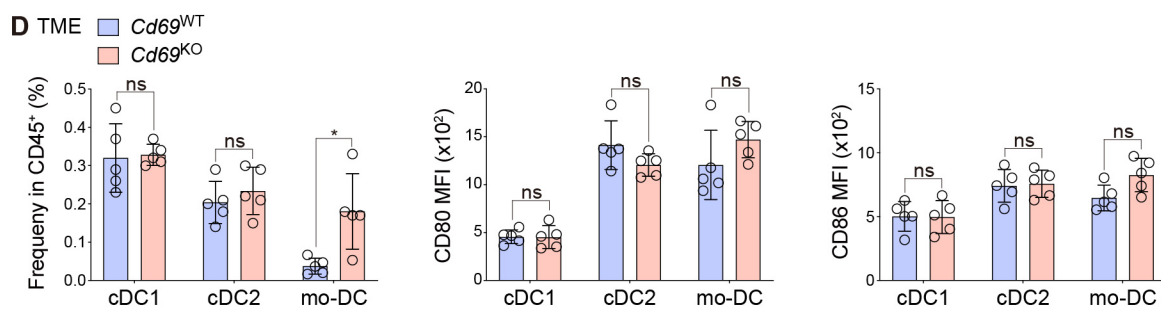
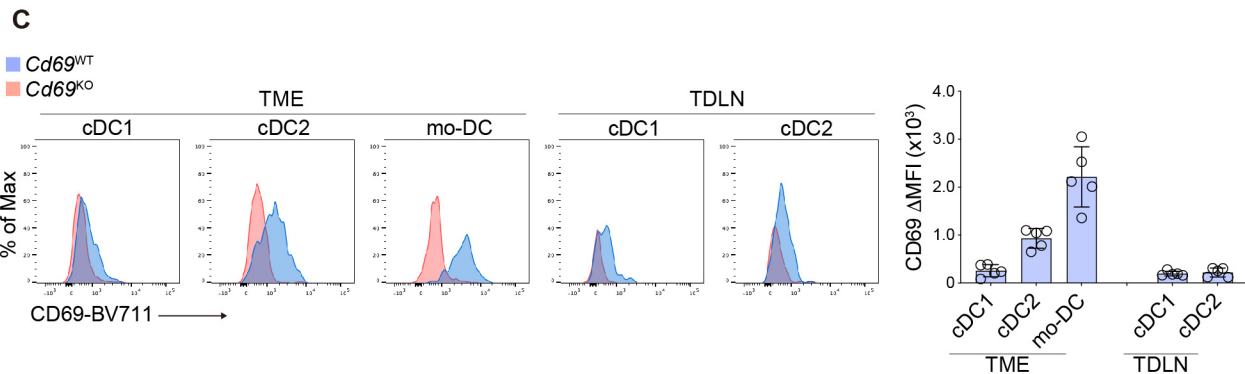
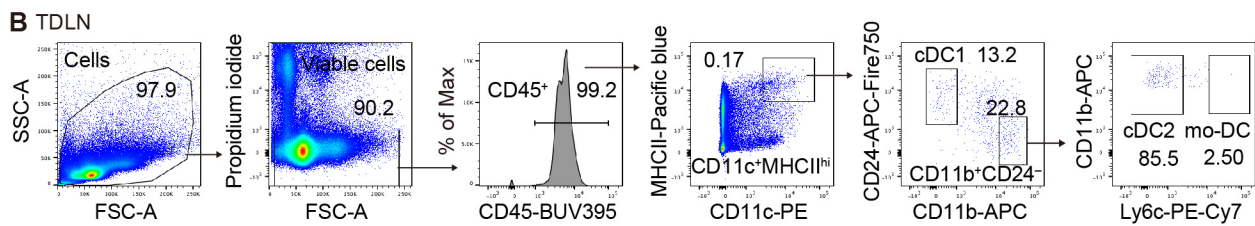
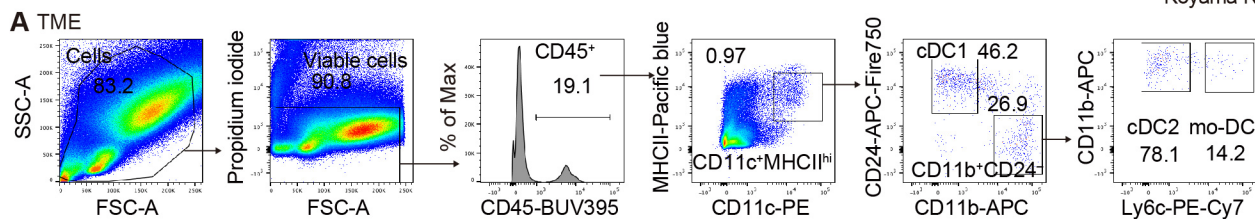
Supplementary Figure S6. Analysis of tumor-specific CD8⁺ T cells in TDLNs. scRNA-seq. **A**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 6-week-old female BALB/c mice. AH1 tetramer⁺ CD8⁺ T cells in TDLNs were isolated at 14 days after inoculation. Experiments were performed in duplicate. Pool of 10 mice in 1 group. A UMAP projection of *Cd69*^{WT} (n=7,014 and 5,807 cells) and *Cd69*^{KO} (n=6,285 and 10,956 cells) cells that were assigned to 5 clusters and color-coded based on the clusters, determined by Seurat. A heatmap of the top 10 genes expressed in each cluster as defined in Fig. 3B. The columns correspond to the cells, the rows correspond to the genes. Cells are grouped by clusters. The color scale is based on a z-score distribution from -2 (blue) to 2 (red). **B**, Single-cell transcript levels of the indicated genes illustrated in the UMAP plots. **C**, Cell-cycle phase (G0/G1, S, or G2/M) of each cell was determined by Seurat and is shown in the UMAP plots. **D**, A schematic outline of CT26 tumor model. **E-F**, CT26 cells were subcutaneously inoculated into 9-week-old female BALB/c mice. AH1 tetramer⁺ CD8⁺ T cells in TDLNs were analyzed at the indicated time. At least n=3 per group. The frequency of CD8⁺ T cell subpopulations in AH1 tetramer⁺PD-1⁺ CD8⁺ T cells (**E**). TOX MFI in AH1 tetramer⁺PD-1⁺ CD8⁺ T cells (**F**). An unpaired two-sided Student's *t*-test was used for the analysis of the flow cytometry data. * P<0.05, ** P<0.01. ns: not significant. Representative data from two experiments with at least three mice are shown. n.d. means not detected.



Supplementary Figure S7. Flow cytometry of OVA-specific CD8⁺ T cells in TDLNs. **A**, A schematic outline of MC38-OVA tumor model. **B**, A representative flow cytometry gating strategy for CD8⁺ T cell subpopulations in TDLNs. MC38-OVA cells were subcutaneously inoculated into 8-week-old C57BL/6 mice. OVA tetramer⁺ CD8⁺ T cells in TDLNs were analyzed at seven days after inoculation. **C-E**, MC38-OVA cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 8-week-old C57BL/6 mice. OVA tetramer⁺ CD8⁺ T cells in TDLNs were analyzed at seven days after inoculation. n=7 per group. The frequency of OVA tetramer⁺ CD8⁺ T cells in CD45⁺ cells. The number indicates the percentage of gated cells. Representative flow plots (left) and a summary of the experiments (right) (**C**). The frequency of CD8⁺ T cell subpopulations in OVA tetramer⁺PD-1⁺ CD8⁺ T cells (**D**). TOX MFI in CD8⁺ T cell subpopulations (**E**). An unpaired two-sided Student's *t*-test was used for the analysis of the flow cytometry data. ** P<0.01, *** P<0.001, **** P<0.0001. Representative data from two experiments with at least five mice are shown (**C-E**).

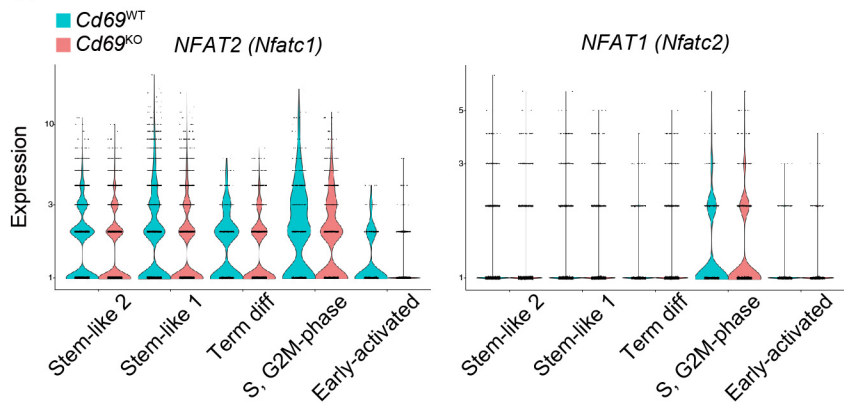


Supplementary Figure S8. The expression of *CD69* and *TOX* in CD8⁺ T cells in human TDLNs. **A**, A UMAP projection of CD8⁺ T cell (CD8 low T, Cytotoxic CD8⁺ T, Naive CD8⁺ T and Exhausted CD8⁺ T) clusters in treatment-naïve human lung adenocarcinoma patients from tumor-containing lung tissue (tLung) and non-metastatic lung-draining lymph nodes (nLN). **B**, The graph represents the percentages of cells from the indicated tissues in each cluster. **C**, The single-cell transcript levels of the indicated genes illustrated in the UMAP plots. **D**, Violin plots showing the expression of the indicated genes in cells from stem-like and term diff clusters in both nLN and tLung as defined in **A**. Each dot represents a single cell. The scRNA-seq dataset was obtained from Kim et al. (35).

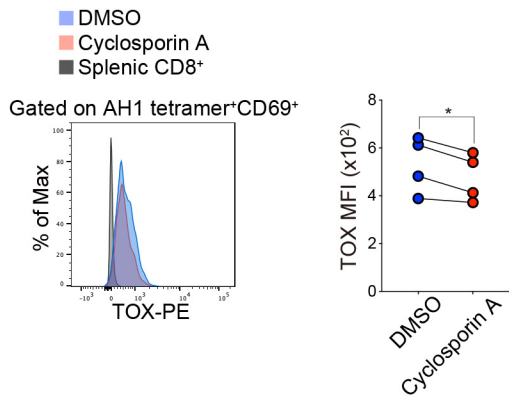


Supplementary Figure S9. Analysis of major dendritic cell populations in the TME and TDLNs. A representative flow cytometry gating strategy for major dendritic cell (DC) populations in the TME (**A**) and TDLNs (**B**). **C**, CT26 cells were subcutaneously inoculated into 12-week-old BALB/c mice. n=5 per group. Delta MFI of CD69 (delta MFI was calculated by subtracting *Cd69*^{KO} MFI from *Cd69*^{WT} MFI) in cDC1, cDC2 and mo-DCs in the TME and cDC1 and cDC2 in TDLNs. **D-E**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 12-week-old female BALB/c mice. n=5 per group. DCs in the TME (**D**) and TDLNs (**E**) were analyzed by flow cytometry. The frequency of DCs in CD45⁺ cells and MFI of the indicated protein were shown. **F-G**, CT26 cells were subcutaneously inoculated into 6-week-old BALB/c mice. n=5 per group. Anti-CD69 Ab or control IgG were intraperitoneally administered to mice twice a week, 3 times. DCs in the TME (**F**) and TDLNs (**G**) were analyzed by flow cytometry. The frequency of DCs in CD45⁺ cells and MFI of the indicated protein were shown. n.d. means not detected. An unpaired two-sided Student's *t*-test was used for the analysis of the flow cytometry data. * P<0.05. not significant. Representative data from two experiments with at least five mice are shown (**C-G**).

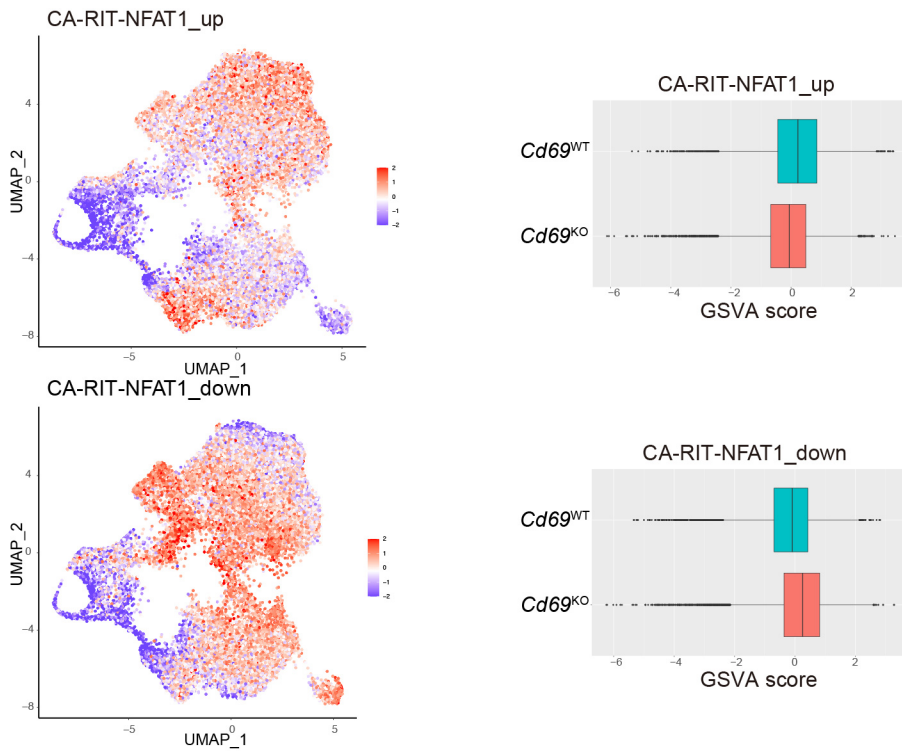
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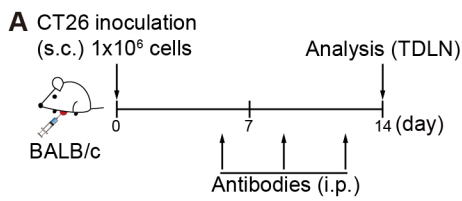
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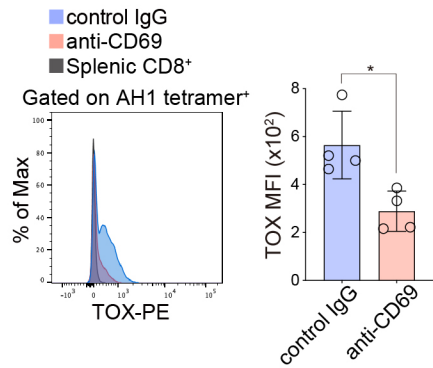
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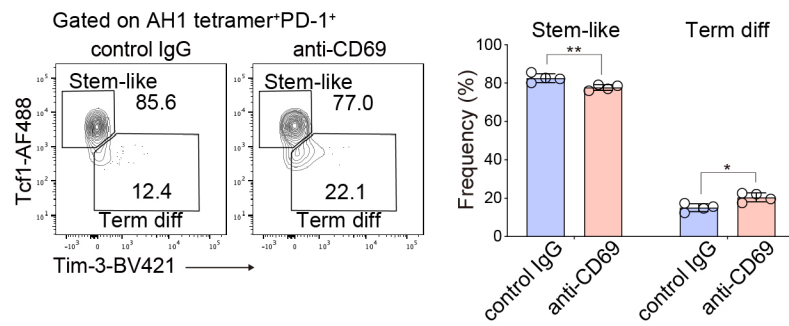
Supplementary Figure S10. CD69 controls NFAT signaling in TDLNs. **A**, Violin plots showing the expression of the indicated genes in cells from each cluster as defined in Fig. 3B. The violin represents the probability density at each value. Each dot represents a single cell. **B**, CT26 cells were subcutaneously inoculated into *Cd69*^{WT} and *Cd69*^{KO} 6-week-old female BALB/c mice. n=4 per group. TDLN cells were cultured *in vitro* in the presence or absence of Cyclosporin A for 4 hours. MFI of TOX in AH1 tetramer⁺CD69⁺ CD8⁺ T cells. **C**, Single-sample gene set variation analysis (ssGSVA) scores of CA-RIT-NFAT1_up (upper) and CA-RIT-NFAT1_down (lower) are projected on UMAPs and box plots. The gene expression profiles of CA-RIT-NFAT1- and mock-transduced CD8⁺ T cells were obtained from Martinez et al.(26). An unpaired two-sided Student's *t*-test was used for the analysis of the flow cytometry data. * P<0.05. Representative data from two experiments with at least four mice are shown (**B**).



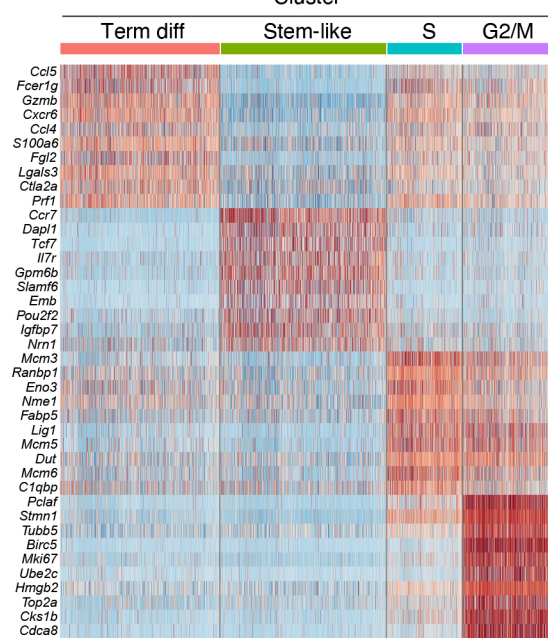
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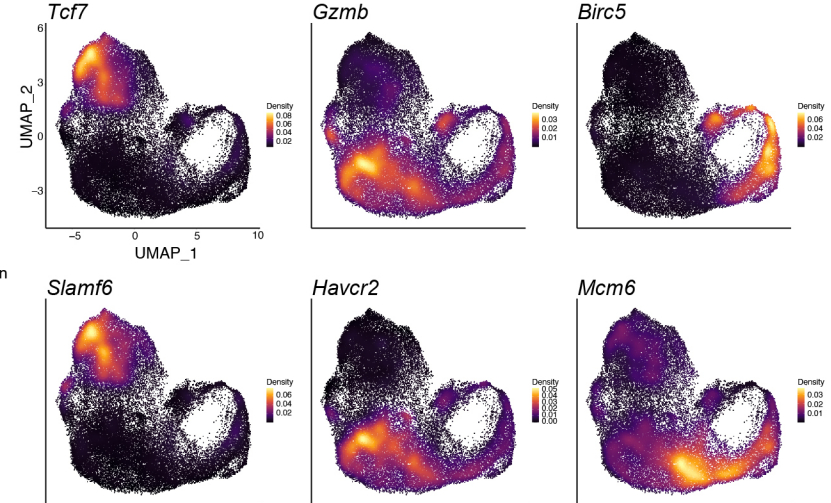
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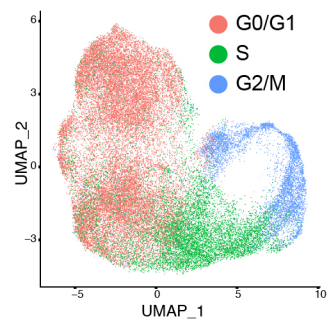
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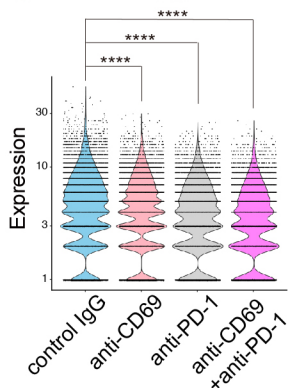
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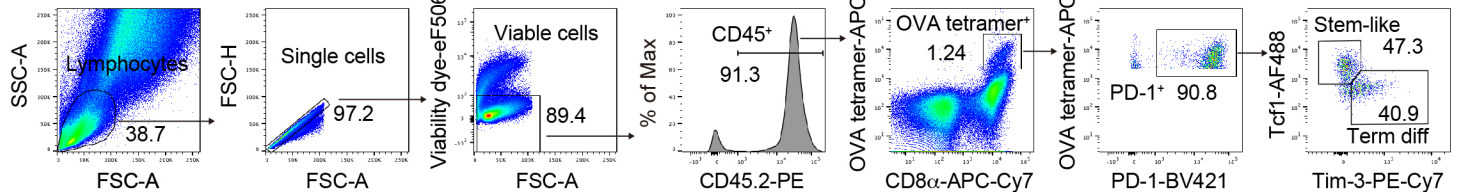
F TME



G TME



H TME



Supplementary Figure S11. Analysis of tumor-specific CD8⁺ T cells from mice treated with therapeutic antibodies. **A**, A schematic outline of the administration of antibodies during CT26 tumor development. CT26 cells were subcutaneously inoculated into 6-week-old female BALB/c mice. Antibodies were intraperitoneally administered to mice twice a week, 3 times. AH1 tetramer⁺ CD8⁺ T cells in TDLNs were analyzed at 14 days after CT26 cell inoculation. n=4 per group. TOX MFI (**B**). The frequency of CD8⁺ T cell subpopulations (**C**). **D-F**, scRNA-seq. CT26 cells were subcutaneously inoculated into 7-week-old female BALB/c mice. Abs were intraperitoneally administered to mice twice a week, 3 times. Intratumoral AH1 tetramer⁺ CD8⁺ T cells were isolated at 14 days after inoculation. Experiments were performed in duplicate. Pool of four mice in 1 group. A UMAP projection of control IgG (n=6,430 and 6,489 cells), anti-CD69 (n=5,933 and 6,213 cells), anti-PD-1 (n=6,706 and 4,950 cells) and anti-CD69 and anti-PD-1 (n=7,094 and 3,674 cells) treated cells that were assigned to 4 clusters and color-coded based on the clusters, determined by Seurat. **D**, A heatmap of the top 10 genes expressed in each cluster as defined in Fig. 5B. The columns correspond to the cells, the rows correspond to the genes. Cells are grouped by clusters. The color scale is based on a z-score distribution from -2 (blue) to 2 (red). **E**, Single-cell transcript levels of the indicated genes illustrated in the UMAP plots. **F**, The cell-cycle phase (G0/G1, S, or G2/M) of each cell was determined by Seurat and is shown in the UMAP plots. **G**, Violin plots showing the expression of the *Tox* gene in cells treated with the indicated antibodies. The violin represents the probability density at each value. Each dot represents a single cell. Statistical test is performed using one-way ANOVA. **H**, A representative flow cytometry gating strategy for intratumoral CD8⁺ T cell subpopulations. B16-OVA cells were subcutaneously inoculated into 7-week-old C57BL/6 mice. Intratumoral OVA tetramer⁺ CD8⁺ T cells were analyzed at 10 days after inoculation. An unpaired two-sided Student's *t*-test was used for the analysis of the flow cytometry data. * P<0.05, ** P<0.01. Representative data from two experiments with at least four mice are shown (**B-C**).